

# A *meso*-Triphenylamine-BODIPY Derivative for the Optical Chemosensing of Metal Ions<sup>†</sup>

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**Abstract:** The design and synthesis of organic molecules for recognition of biologically/environmentally important metal ions has emerged as a highly regarded research field. The BODIPY (4,4-difluoro-4-bora-3a,4a-diaza-s-indacene) core is a versatile signaling molecule that can be fine-tuned with functional groups to create selective binding sites to improve its optical properties. As an extension of the work developed in our research group, we report the synthesis and characterization of a BODIPY functionalized with triphenylamino and a formyl group at the *meso* and 2-position, respectively, for the highly selective detection of Cu<sup>2+</sup> and Fe<sup>3+</sup>. The preliminary study of the BODIPY derivative as optical chemosensor was carried out in acetonitrile solution in the presence of different cations, and interactions with Cu<sup>2+</sup> and Fe<sup>3+</sup> induced a perceptible color change. UV-visible titrations showed changes in the absorption spectra upon the addition of three equivalents of each cation, with the appearance of a new absorption band at 693 nm.

**Keywords:** BODIPY; metal ions; optical chemosensor; synthesis

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## 1. Introduction

Great efforts have been devoted to developing organic molecules as optical chemosensors for application in fields of biomolecular analysis, medical diagnosis, and environmental monitoring. In particular, metal ions have become a prime target, considering that they are essential elements for biological systems but in uncontrolled amounts may represent a critical issue. Classical chemosensors for metal ions contain molecular structures based on polyamines, polyethers, polysulfides, carboxylic acids, hydroxamic acids, and open-chain or macrocyclic structures [1–4].

In the last two decades, BODIPY derivatives have emerged as a novel class of chemosensors for molecular recognition and biological fluorescent labelling. BODIPY shows remarkable optical properties, such as sharp absorption and emission patterns, high molar extinction coefficient of absorbance, high fluorescence quantum yield, and good photostability under physiological conditions [5–7].

The functionalization of the BODIPY framework enables the introduction of highly selective/sensitive binding sites and, simultaneously, modulates its photophysical properties. BODIPYs modified at the *meso*-position show greater stability than their *meso*-unsubstituted analogues. In fact, diverse aldehydes can be used to prepare *meso*-substituted derivatives which allows the design of innovative compounds bearing a large range of functional groups. Furthermore, the BODIPY core is inherently an electron-rich heteroaromatic structure and is therefore very susceptible to regioselective electrophilic aromatic substitution reactions, preferably at positions 2 and 6 [8,9].

In continuation of our research group's work in the field of chromofluorogenic chemosensors [10–12], we report on the synthesis, characterization, and evaluation of a

*meso*-triphenylamine-BODIPY derivative as optical chemosensor of Cu<sup>2+</sup> and Fe<sup>3+</sup> in acetonitrile solution.

## 2. Experimental Section

### 2.1. Methods and Materials

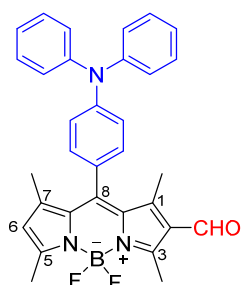
Nuclear Magnetic Resonance (NMR) spectra were obtained on a Bruker Avance III 400 at an operating frequency of 400 MHz, using the solvent peak as an internal reference. The solvents are indicated in parenthesis before the chemical shifts values ( $\delta$  relative to tetramethylsilane (TMS)). Mass spectrometry analyses were performed at the “C.A.C.T.I.-Unidad de Espectrometria de Masas” at the University of Vigo, Spain. All reagents were purchased from Sigma-Aldrich, Acros, and Fluka and used as received. BODIPY precursor 1 was synthesized as previously reported [13]. Thin Layer Chromatography TLC analyses were carried out on 0.25 mm thick precoated silica plates (Merck Fertigplatten Kieselgel 60F254) and spots were visualized under UV light. Chromatography on silica gel was carried out on Merck Kieselgel (230–400 mesh). UV-visible absorption spectra were obtained using a Shimadzu UV/2501PC spectrophotometer. Fluorescence spectra were collected using a Horiba FluoroMax-4 spectrofluorometer. The relative fluorescence quantum yield was determined by using a  $1.0 \times 10^{-5}$  M solution of rhodamine 6G in ethanol as standard ( $\Phi_F = 0.95$ ) [14,15].

### 2.2. Synthesis of BODIPY Derivative 2

A mixture of *N,N*-dimethylformamide (23 mmol) and POCl<sub>3</sub> (18.2 mmol) was stirred for 5 min at 0 °C under N<sub>2</sub> atmosphere. The mixture was allowed to reach room temperature and stirred for an additional 30 min. BODIPY precursor 1 (0.127 mmol) dissolved in dichloroethane (7 mL) was added dropwise while stirring. The reaction mixture was heated for 2 h at 50 °C. After cooling, the solution was poured slowly into 40 mL of saturated sodium bicarbonate aqueous solution at 0 °C and stirred during 30 min at room temperature. Ethyl acetate (5 mL) was added to the reaction mixture and the resulting organic layer was separated and washed with water (2 × 50 mL). The organic layer was dried with anhydrous MgSO<sub>4</sub>, filtered, and the solvent was evaporated. The crude residue was purified by a silica gel chromatography column, using dichloromethane as eluent. The BODIPY derivative 2 (Figure 1) was obtained as a dark red solid (6.8 mg, 15%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.66 (s, 3H, CH<sub>3</sub>-7), 1.89 (s, 3H, CH<sub>3</sub>-1), 2.62 (s, 3H, CH<sub>3</sub>-5), 2.83 (s, 3H, CH<sub>3</sub>-3), 6.15 (s, 1H, H-6), 7.09–7.21 (m, 10H, Ar-H), 7.30–7.34 (m, 4H, Ar-H), 10.05 (s, 1H, CHO) ppm.

MS (ESI) *m/z* (%): 521 ((M + 2)<sup>+</sup>, 35), 520 ((M + 1)<sup>+</sup>, 100), 519 ((M)<sup>+</sup>, 34), 453 (7), 232 (2); HRMS (ESI) *m/z*: (M + 1)<sup>+</sup> calculated for C<sub>32</sub>H<sub>29</sub>BF<sub>2</sub>N<sub>3</sub>O, 520.2366; found 520.2381.



**Figure 1.** Structure of BODIPY derivative 2.

### 2.3. Chemosensing Study of BODIPY Derivative 2 and UV-Visible Absorption Titrations

The evaluation of BODIPY derivative 2 as an optical chemosensor was carried out in the presence of several cations (Ag<sup>+</sup>, K<sup>+</sup>, Li<sup>+</sup>, Pb<sup>2+</sup>, Mn<sup>2+</sup>, Cd<sup>2+</sup>, Cu<sup>2+</sup>, Co<sup>2+</sup>, Pd<sup>2+</sup>, Ni<sup>2+</sup>, Ca<sup>2+</sup>, Hg<sup>2+</sup>, Zn<sup>2+</sup>, Fe<sup>2+</sup>, Fe<sup>3+</sup>, and Al<sup>3+</sup>) with environmental and biomedical relevance. Solutions of

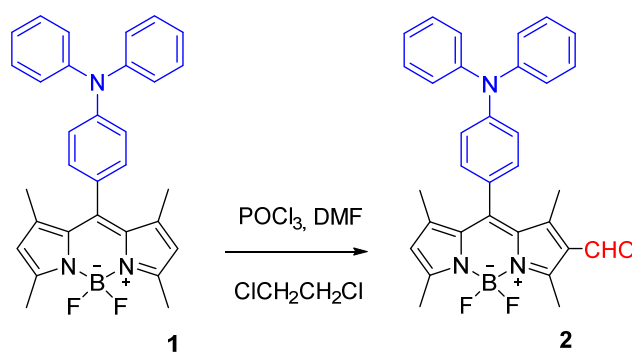
the BODIPY derivative and cations were prepared in acetonitrile at  $1.0 \times 10^{-5}$  M and  $1.0 \times 10^{-2}$  M, respectively.

A preliminary study was carried out *via* the addition of 50 equivalents of each cation to the solution of the BODIPY derivative. UV-visible absorption titration experiments were performed by the sequential addition of  $\text{Cu}^{2+}$  and  $\text{Fe}^{3+}$ , as a result of the previous study which revealed selectivity towards these cations. The stability constants were determined from UV-visible spectroscopic titration data, using the HypSpec software [16].

### 3. Results and Discussion

#### 3.1. Synthesis of BODIPY Derivative 2

BODIPY precursor **1** was prepared according to a procedure described previously [13]. The synthesis of BODIPY derivative **2** was obtained through the Vilsmeier–Haack formylation of BODIPY precursor **1**, *meso*-substituted with a triphenylamino group, using *N,N*-dimethylformamide (DMF) and phosphorylchloride ( $\text{POCl}_3$ ) in dichloroethane (Scheme 1). The pure compound was obtained as a dark red solid in 15% yield.



**Scheme 1.** The synthesis of BODIPY derivative **2**.

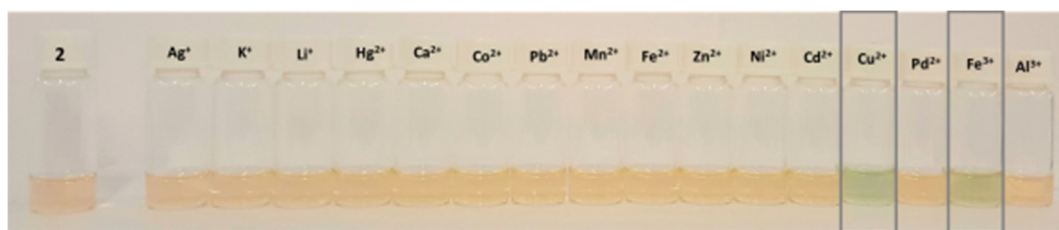
The presence of a formyl group at position 2 of the BODIPY core was confirmed by  $^1\text{H}$  NMR spectroscopy, with the appearance of a singlet at  $\delta$  10.05 ppm. Additionally, the obtained data from mass spectrometry was in agreement with the expected structure.

#### 3.2. Photophysical Characterization of BODIPY Derivative 2

The photophysical properties of BODIPY derivative **2** were investigated in acetonitrile solution. The compound showed an intense absorption band ( $\log \epsilon = 4.02$ ) at 491 nm. Upon excitation at 460 nm, the compound exhibited an emission band at 515 nm. The relative fluorescence quantum yield, determined by using rhodamine 6G in ethanol as standard ( $\Phi_F = 0.95$ ), was found to be low ( $\Phi_F = 0.010$ ). The small quantum yield measured could be ascribed to a carbonyl electron-withdrawing effect exerted by the formyl group linked to the BODIPY core [17].

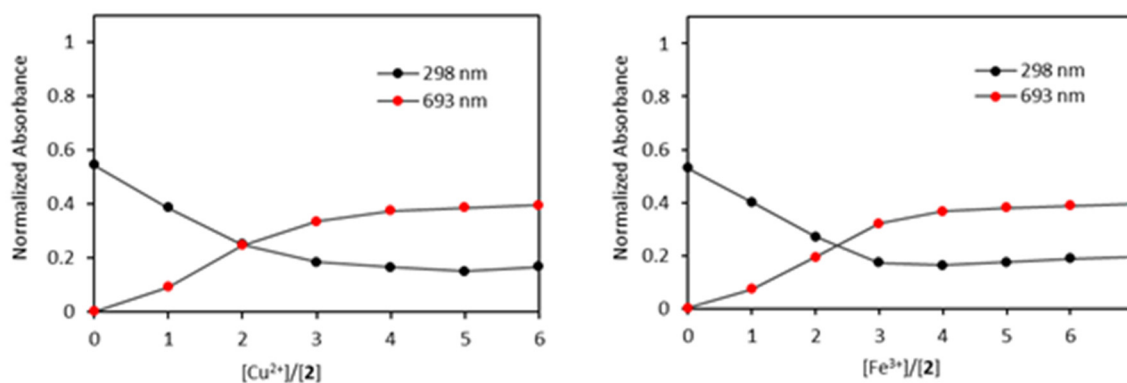
#### 3.3. Chemosensing Studies of BODIPY Derivative 2 and Spectrophotometric Titration

A preliminary evaluation of BODIPY derivative **2** as an optical chemosensor was carried out in the presence of several cations in acetonitrile solution. The chromogenic response of the BODIPY derivative was visible to the naked eye in the presence of  $\text{Cu}^{2+}$  and  $\text{Fe}^{3+}$ . In Figure 2 the color modulation upon addition of 50 equivalents of each cation is observed. As shown, a moderate color change from orange to blue-green exclusively occurred upon the addition of  $\text{Cu}^{2+}$  and  $\text{Fe}^{3+}$ , while other cations were unable to induce a perceptible color change.



**Figure 2.** Color changes observed for BODIPY derivative **2** ( $1.0 \times 10^{-5}$  M in acetonitrile) upon the addition of 50 equivalents of different cations.

The UV-visible absorption titration of BODIPY derivative **2** was performed in acetonitrile with  $\text{Cu}^{2+}$  and  $\text{Fe}^{3+}$ , as a result of the previous study which revealed a selective chromogenic response towards these cations. As shown in Figure 3, a similar behaviour was observed upon the addition of increasing quantities of  $\text{Cu}^{2+}$  and  $\text{Fe}^{3+}$ . The intensity of the absorption band at 298 nm decreased, while a new red-shifted band appeared at 693 nm with a maximum absorbance reached at approximately three equivalents of each cation.



**Figure 3.** Absorbance at 298/693 nm as a function of the number of equivalents of each cation. Data was obtained from the UV-visible titration spectra of BODIPY derivative **2** ( $1.0 \times 10^{-5}$  M in acetonitrile) upon the addition of increasing quantities of  $\text{Cu}^{2+}$  (left) and  $\text{Fe}^{3+}$  (right).

Moreover, the interaction between BODIPY derivative **2** with these cations was studied by the determination of the stability constants from the UV-visible spectroscopic titration data, using the HypSpec software. The results suggested the formation of 2:1 cation–BODIPY stoichiometry complexes with a logarithm of the stability constant ( $\log K$ ) of 10.61 for  $\text{Cu}^{2+}$  and 10.19 for  $\text{Fe}^{3+}$ .

#### 4. Conclusions

BODIPY derivative **2** functionalized with triphenylamino and a formyl group at *meso* and 2-position, respectively, was synthesized by Vilsmeier–Haack formylation of the BODIPY precursor **1** and its molecular structure was confirmed by  $^1\text{H}$  NMR and mass spectrometry. The recognition of both  $\text{Cu}^{2+}$  and  $\text{Fe}^{3+}$  in acetonitrile solution among several cations was observed through a perceptible color change from orange to blue-green. UV-visible titrations showed alterations in the absorption spectra upon the addition of three equivalents of each cation with the appearance of a new absorption band at 693 nm, which clearly indicates that probe **2** can be used for detection of  $\text{Cu}^{2+}$  and  $\text{Fe}^{3+}$  ions in solution.

**Author Contributions:** Conceptualization: M.M.M.R.; Experimental work: R.C.R.G., S.C.S.P.; Methodology and formal analysis: S.P.G.C., M.M.M.R.; Writing of original draft: R.C.R.G., S.C.S.P., S.P.C.C., M.M.M.R.; Review and editing: S.P.C.C., M.M.M.R. All authors have read and agreed to the published version of the manuscript.

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**Conflicts of Interest:** The authors declare no conflicts of interest.

## References

1. Wang, B.; Anslyn, E.V. *Chemosensors: Principles, Strategies and Applications*, 1st ed.; John Wiley & Sons: Hoboken, NJ, USA, 2011; Volume 15, pp. 227–295.
2. You, L.; Zha, D.; Anslyn, E.V. Recent Advances in Supramolecular Analytical Chemistry Using Optical Sensing. *Chem. Rev.* **2015**, *115*, 7840–7892, doi:10.1021/cr5005524.
3. Kaur, B.; Kaur, N.; Kumar, S. Colorimetric Metal Ion Sensors—A Comprehensive Review of the Years 2011–2016. *Coord. Chem. Rev.* **2018**, *358*, 13–69, doi:10.1016/j.ccr.2017.12.002.
4. Formica, M.; Fusi, V.; Giorgi, L.; Micheloni, M. New fluorescent chemosensors for metal ions in solution. *Coord. Chem. Rev.* **2012**, *256*, 170–192, doi:10.1016/j.ccr.2011.09.010.
5. Loudet, A.; Burgess, K. BODIPY Dyes and their Derivatives: Syntheses and Spectroscopic Properties. *Chem. Rev.* **2007**, *107*, 4891–4932, doi:10.1021/cr078381n.
6. Boens, N.; Verbelen, B.; Dehaen, W. Postfunctionalization of the BODIPY Core: Synthesis and Spectroscopy. *Eur. J. Org. Chem.* **2015**, *2015*, 6577–6595, doi:10.1002/ejoc.201500682.
7. Lu, H.; Shen, Z. *BODIPYs and Their Derivatives: The Past, Present and Future*; Frontiers Media SA: Lausanne, Switzerland, 2020; doi:10.3389/978-2-88963-786-7.
8. Bañuelos, J.; Arroyo-Córdoba, I.J.; Valois-Escamilla, I.; Alvarez-Hernández, A.; Peña-Cabrera, E.; Hu, R.; Ben, Z.T.; Esnal, I.; Martínez, V.; López Arbeloa, I. Modulation of the Photophysical Properties of BODIPY Dyes by Substitution at Their *Meso* Position. *RSC Adv.* **2011**, *1*, 677, doi:10.1039/c1ra00020a.
9. Zhu, S.; Bi, J.; Vegesna, G.; Zhang, J.; Luo, F.-T.; Valenzano, L.; Liu, H. Functionalization of BODIPY Dyes at 2,6-Positions through Formyl Groups. *RSC Adv.* **2013**, *3*, 4793, doi:10.1039/c3ra22610g.
10. Okda, H.E.; El Sayed, S.; Ferreira, R.C.M.; Gonçalves, R.C.R.; Costa, S.P.G.; Raposo, M.M.M.; Martínez-Mañez, R.; Sancenón, F. *N,N*-Diphenylanilino-heterocyclic aldehyde-based chemosensors for UV-vis/NIR and fluorescence Cu(II) detection. *New J. Chem.* **2019**, *43*, 7393–7402, doi:10.1039/c9nj00880b.
11. Presti, M.L.; Martínez-Mañez, R.; Ros-Lis, J.V.; Batista, R.M.F.; Costa, S.P.G.; Raposo, M.M.M.; Sancenón, F. A Dual Channel Sulphur-containing Macrocyclic Functionalised BODIPY Probe for the Detection of Hg(II) in Mixed Aqueous Solution. *New J. Chem.* **2018**, *42*, 7863–7868, doi:10.1039/c7nj04699e.
12. Esteves, C.I.C.; Ferreira, R.C.M.; Raposo, M.M.M.; Costa, S.P.G. New Fluoroionophores for Metal Cations Based on Benzo[*d*]Oxazol-5-yl-Alanine Bearing Pyrrole and Imidazole. *Dyes Pigments* **2018**, *151*, 211–218, doi:10.1016/j.dyepig.2017.12.040.
13. Gonçalves, R.C.R.; Nogueira, M.B.; Costa, S.P.G.; Raposo, M.M.M. Functionalized BODIPY Derivatives as Potential Fluorescent Labels. *Proceedings* **2019**, *9*, 36, doi:10.3390/ecsoc-22-05701.
14. Montalti, M.; Credi, A.; Prodi, L.; Gandolfi, M.T. *Handbook of Photochemistry*, 3rd ed.; CRC Press: Boca Raton, FL, USA, 2006.
15. Demas, J.N.; Crosby, G.A. Measurement of photoluminescence quantum yields. *Rev. J. Phys. Chem.* **1971**, *75*, 991–1024, doi:10.1021/j100678a001.
16. Gans, P.; Sabatini, A.; Vacca, A. Investigation of equilibria in solution. Determination of equilibrium constants with the HYPERQUAD suite of programs. *Talanta* **1996**, *43*, 1739–1753, doi:10.1016/0039-9140(96)01958-3.
17. Valeur, B.; Berberan-Santos, M.N. *Molecular Fluorescence: Principles and Applications*; Wiley-VCH: Weinheim, Germany, 2012.