Universidade do Minho Escola de Medicina

Diana Marina Fonseca Rodrigues

Characterization of sex-specific nociceptive, emotional and electrophysiological impairments
after the induction of the chronic constriction injury model of experimental neuropathic pain Diana Marina Fonseca Rodrigues **Characterization of sex-specific nociceptive, emotional and electrophysiological impairments**
Diana Marina Fonseca Rodrigues after the induction of the chronic constriction injury model of e Diana Marina Fonseca Rodrigues

 $\frac{1}{2}$

 $\frac{1}{4}$

- Characterization of sex-specific
- nociceptive, emotional and
- electrophysiological impairments after the
- induction of the chronic constriction injury
- model of experimental neuropathic pain

Universidade do Minho Escola de Medicina

Diana Marina Fonseca Rodrigues

Characterization of sex-specific nociceptive, emotional and electrophysiological impairments after the induction of the chronic constriction injury model of experimental neuropathic pain

Dissertação de Mestrado Mestrado em Ciências da Saúde

Trabalho efetuado sob a orientação da Professora Filipa Pinto-Ribeiro e da Doutora Diana Amorim

DIREITOS DE AUTOR E CONDIÇÕES DE UTILIZAÇÃO DO TRABALHO POR TERCEIROS

Este é um trabalho académico que pode ser utilizado por terceiros desde que respeitadas as regras e boas práticas internacionalmente aceites, no que concerne aos direitos de autor e direitos conexos.

Assim, o presente trabalho pode ser utilizado nos termos previstos na licença abaixo indicada.

Caso o utilizador necessite de permissão para poder fazer um uso do trabalho em condições não previstas no licenciamento indicado, deverá contactar o autor, através do RepositoriUM da Universidade do Minho.

Licença concedida aos utilizadores deste trabalho

Atribulção-NãoComercial-SemDerivações **CC BY-NC-ND**

https://creativecommons.org/licenses/by-nc-nd/4.0/

AGRADECIMENTOS

Gostaria de apresentar o meu mais sincero obrigada a todas as pessoas que, de alguma forma, contribuíram para a realização deste trabalho, e sem os quais não seria possível.

À professora Filipa Pinto-Ribeiro, pelo apoio, amizade e muita paciência. Pela confiança, por todos os conselhos, e por me ter guiado durante estes anos, pessoal e profissionalmente. Por todas as gargalhadas e a sempre boa disposição, muito obrigada.

À Diana Amorim, pela amizade, ajuda e por me ensinares tudo o que sei. Obrigada por todo o apoio e por estares sempre disponível a ajudar-me.

Ao professor Armando Almeida, pela oportunidade de desenvolver este trabalho.

Ao meu grupo, em especial à Inês Laranjeira e ao João Barbosa. Pela vossa amizade e porque sem a vossa ajuda, disponibilidade e boa disposição nada disto era possível.

A todos os NeRDs e ao staff do ICVS.

Aos meus amigos. Aos que no Porto ficaram e que a distância não separou. Em especial, um muito obrigada Inês, Joana, Helena e João Viana - graças à vossa amizade Braga ganhou outro significado para mim.

Ao João, por seres o meu suporte e continuares comigo todos estes anos.

Aos meus pilares, mãe, pai e irmã. Obrigada aos meus pais pela luta diária, por tudo o que sacrificaram por mim, pelo apoio incondicional e imbatível. Não estaria aqui se não fossem vocês e tudo o que sou devo-o a vós. E à minha irmã que, por mais chatinha que sejas, sei que posso sempre contar contigo.

Este trabalho foi realizado no Instituto de Investigação em Ciências da Vida e da Saúde (ICVS) da Universidade do Minho. O financiamento provém do projeto NORTE-01-0145-FEDER-000013, do Programa Operacional Regional do Norte (NORTE 2020/FEDER), e do projeto POCI-01-0145- FEDER-007038, sobre o Programa Operacional Fatores de Competitividade (COMPETE) e de fundos nacionais da Fundação para a Ciência e Tecnologia (FCT).

STATEMENT OF INTEGRITY

I hereby declare having conducted this academic work with integrity. I confirm that I have not used plagiarism or any form of undue use of information or falsification of results along the process leading to its elaboration.

I further declare that I have fully acknowledged the Code of Ethical Conduct of the University of Minho.

RESUMO

Caracterização nociceptiva, emocional e eletrofisiológica do modelo experimental de dor neuropática por constrição crónica em fêmeas Wistar Han

A dor crónica neuropática afeta 7-10% da população e é frequentemente acompanhada por distúrbios emocionais, nomeadamente ansiedade e depressão. Esta patologia reduz significativamente a qualidade de vida dos pacientes, afetando as suas capacidades físicas, cognitivas, emocionais e sociais e, consequentemente, interferindo com o tratamento. No entanto, apesar de a dor crónica e dos distúrbios emocionais que lhe estão associados serem mais prevalentes em mulheres, poucos trabalhos são realizados em animais fêmea. Não obstante o modelo de lesão por constrição crónica (CCI) mimetizar neuropatias humanas em termos de hiperalgesia mecânica e térmica, alodinia e dor espontânea em ambos os sexos, são escassos os dados sobre distúrbios emocionais em ratos fêmea.

Neste trabalho, ratos jovens (Wistar Han) foram divididos em cinco grupos: machos com CCI (CCI.), fêmeas gonadicamente intactas (SHAM/SHAM), fêmeas ovariectomizadas (SHAM/OVX), fêmeas gonadicamente intactas com CCI (CCI/SHAM) e fêmeas ovariectomizadas com CCI (CCI/OVX). A avaliação nocicetiva iniciou-se na semana anterior à cirurgia de CCI, perdurando 5 semanas, após as quais o comportamento anxiodepressivo foi também avaliado. No período pós-operatório, os animais com CCI apresentaram alterações na marcha, e o desenvolvimento de alodinia mecânica e térmica ao frio. Todas as fêmeas CCI desenvolveram alodinia mecânica e este comportamento instalou-se mais cedo nas CCI/OVX comparativamente às CCI/SHAM. A indução de CCI em fêmeas não levou ao desenvolvimento de comportamentos ansiosos e depressivos, embora a ovariectomia tenha induzido um comportamento tipo-anedónico, independentemente de CCI. As gravações eletrofisiológicas *single cell* na medula rostral ventromedial (RVM), área mediadora do processamento nociceptivo, sugerem um aumento da pronociceção descendente após neuropatia crónica. Estes resultados demonstram a existência de diferenças entre sexos na comorbidade entre a dor neuropática e distúrbios emocionais no modelo de CCI, modulada pelo controlo descendente da RVM.

Palavras-chave: ansiedade; depressão; dor neuropática; eletrofisiologia; histologia; lesão por constrição crónica; medula rostral ventromedial (RVM); nervo ciático.

ABSTRACT

Characterization of sex-specific nociceptive, emotional and electrophysiological impairments after the induction of the chronic constriction injury model of experimental neuropathic pain

Chronic neuropathic pain affects 7-10% of the population and is often accompanied by comorbid emotional impairments that adversely affect nociceptive symptomatology. This condition greatly reduces the quality of life of the patients, impairing physical, cognitive, emotional and social functioning which, consequently, interferes with treatment. Importantly, although chronic pain and emotional disorders are more prevalent in women, only a few publications focus on female animals. While the chronic constriction injury (CCI) model has been shown to mimic human neuropathies regarding mechanical/thermal hyperalgesia, allodynia and spontaneous pain in both sexes, data on CCI-induced emotional impairments on female rats remains scarce.

In this work, young adult rats (Wistar Han) were divided into five groups: males with CCI (CCI_M), gonadally intact females (SHAM/SHAM), ovariectomized females (SHAM/OVX), ovariectomized females with CCI (CCI/OVX) and gonadally intact females with CCI (CCI/SHAM). Nociceptive testing began before CCI surgery and was performed weekly throughout 5 weeks after which the anxiodepressive-like behaviour was also assessed. In the postoperative period, CCI animals displayed visible gait abnormalities, limping and guarding the affected hind paw, and the development of mechanical and cold allodynia from week 1 onwards. CCI developed mechanical allodynia, with CCI/OVX animals displaying symptoms prior to CCI/SHAM females. While no differences were found between CCI and SHAM animals concerning anxietyand depressive-like behaviours, ovariectomy induced anhedonic-like behaviour, regardless of CCI surgery. Single cell electrophysiological data from the rostral ventromedial medulla (RVM), an area mediating nociceptive processing, suggest an increase in descending pronociception after chronic neuropathy. These results demonstrate the existence of sex differences on the comorbidity between neuropathic pain on the CCI model, modulated by RVM descending control.

Keywords: anxiety; chronic constriction injury; depression; electrophysiology; histology; neuropathic pain; rostral ventromedial medulla (RVM); sciatic nerve

TABLE OF CONTENTS

ABBREVIATIONS

- ACC anterior cingulate cortex
- AMY amygdala
- ANOVA analysis of variance
- CCI chronic constriction injury
- CVLM caudal ventrolateral medulla
- **DMH** dorsomedial nucleus of the hypothalamus
- **DRt** dorsal reticular nucleus
- DRG dorsal root ganglia
- **EPM** elevated plus maze test
- FST forced swimming test
- Gi gigantocellularis reticularis
- GiA gigantocellularis pars alpha
- IASP International Association for the Study of Pain
- **LDB** light/dark box test
- LPGi nucleus paragigantocellularis lateralis
- MB marble burying test
- NSFT novelty-supressed feeding test
- NS nociceptive-specific neurons
- OVX ovariectomy
- OF open field test
- PB parabrachial area
- PAG periaqueductal grey
- PEAP place escape/avoidance test
- PFC prefrontal cortex
- **PNC** partial nociceptive convergent neurons
- **PWT** paw withdrawal threshold
- RMg nucleus raphe magnus
- RVM rostral ventromedial medulla
- SDH spinal dorsal horn
- **SEM** standard error of the mean
- **SNI** spared nerve ligation
- **SNL** spinal nerve ligation
- SPT sucrose preference test
- **TNC** total nociceptive convergent neurons
- **TST** tail suspension test
- WDR wide-dynamic range neurons

LIST OF FIGURES

LIST OF TABLES

1.1. Nociception

Pain is defined by the International Association for the Study of Pain (IASP, 2018) as an unpleasant sensory and emotional experience associated with actual or potential tissue damage or described in terms of such damage. It is thus a multidimensional experience, comprising sensorial, emotional-affective and cognitive components and whose treatment remains a challenge¹. Pain is a fundamental physiological process that occurs continuously, protecting the organism from potential or actual injury and/or damage. In basic research, it is, however, important to distinguish between pain and nociception, as the latter refers only to the process by which noxious stimuli are encoded and processed².

1.2. Nociceptive transmission

Nociception is encoded and transmitted to the spinal cord by primary afferent neurons³. There are three major classes of primary afferents: Aβ, Aδ, and C-fibers₄, each with different properties, which allow them to respond to and transmit different types of information. Aβ-fibres are large myelinated fibres (4-8 µm diameter), fast conducting of innocuous mechanical stimuli (up to 70 m.s¹)⁵, and have low activation thresholds. Aδ-fibres are smaller thinly myelinated fibres (2-6 µm diameter), slower-conducting than Aβfibres (about 20 m.s¹) and mediate the acute and well-localized "first pain"⁶. Lastly, C-fibres are thin unmyelinated fibres (0.4-1.2 µm diameter), being the slowest conducting fibres (less than 2 m.s¹)^{4,6,7} and are responsible for the "second pain"⁶.

Since Aδ and C-fibres respond to noxious stimuli (mechanical, thermal or chemical) they are also designated as nociceptors⁷. The cell bodies of these fibres are located in the dorsal root ganglia (DRG) or the trigeminal ganglion and have a small axonal branch projecting to the spinal cord and a long branch extending to the periphery⁸. Primary afferents synapse in the dorsal horn of the spinal cord (SDH, Spinal Dorsal Horn) in an organized manner with most of Aδ and C-fibres terminating mainly in laminae I-II, and only a few reaching deeper laminae (Figure 1).

In the dorsal horn, there are two types of neurons, regarding their stimulus-response properties. In the superficial SDH (laminae I-II), we mainly find nociceptive-specific (NS) neurons, which only respond to painful stimuli and receive inputs from Aδ- and C-fibers⁶. The other type of neurons is the wide dynamic range (WDR) neurons, which can be found mainly in laminae III-VI. WDR neurons receive input from all types of peripheral fibres, responding to both innocuous and noxious stimuli⁴.

Figure 1. Summary of pain processing pathways. A peripheral noxious stimulus is signalled via specialized primary afferent fibres, called nociceptors. This nociceptive information is then conducted to the dorsal horn of the spinal cord and then transmitted via ascending nociceptive pathways, through areas such as the parabrachial area (PB), periaqueductal grey (PAG) and the thalamus. This information is processed according to the sensory-discriminative, motivational-affective and cognitive-evaluative dimensions of pain (perception). Descending pathways modulate nociceptive transmission, acting either by promoting or suppressing nociception. Limbic areas such as the limbic anterior cingulate and insular cortex, amygdala and the hypothalamus, project to the PAG, which then modulates descending pain transmission from the afferent pain system indirectly through the rostral ventromedial medulla (RVM) in the brainstem. Adapted⁶.

1.3. Ascending pathways and supraspinal processing

According to their axonal projections, the SDH is comprised by two main types of neurons: (i) interneurons, with short axons, that communicate locally, and ii) projection neurons, with long axons, that communicate supraspinally, conveying nociceptive inputs for further processing⁹. Nociceptive inputs are conveyed through five major ascending pathways¹⁰ ([Figure 2](#page-17-1)).

The pathways responsible for the discriminative component of pain, signalling the location, intensity, and quality of the stimulus, target mainly somatosensory cortical areas. Those targeting additional cortical and brainstem pathways are related to the affective-motivational component of pain (the unpleasantness of the pain experience).

The main ascending pathways are:

- (i) The spinothalamic tract transmits nociceptive information to the thalamus, which is then relayed to cortical structures, such as the somatosensory areas S1 and S2, the insular cortex, the anterior cingulate cortex, and the medial prefrontal cortex (mPFC)¹¹ (**Figure 2A**). Through this pathway, there is an integration of the sensory-discriminative (lateral spinothalamic tract) and motivationalaffective (medial spinothalamic tract) components of the pain experience¹²⁻¹⁵.
- (ii) The spinoreticular tract projects to the brainstem reticular formation, terminating in the thalamus¹³ (Figure 2B).
- (iii) The spinomesencephalic tract targets brainstem nuclei such as the locus coeruleus and the periaqueductal grey (PAG), a major antinociceptive region⁴ (Figure 2C). The PAG is in close interaction with limbic structures, such as the amygdala (AMY) ¹⁶–¹⁸ and the prefrontal cortex (PFC)¹⁹ .
- (iv) In the cervicothalamic tract, the fibres ascend in the medial lemniscus and terminate in the ventral posterolateral nucleus of the thalamus.
- (v) The spinohypothalamic tract projects mainly to the hypothalamus, but also to autonomic control centres in the thalamus and AMY²⁰.

1.4. Descending modulation

After supraspinal processing, nociception is modulated by descending pathways that can either facilitate (pronociception) or inhibit (antinociception) nociceptive transmission in the SDH6,21 . Several supraspinal nuclei are recognized as involved in descending modulation, including the dorsal reticular nucleus (DRt)²², the caudal ventrolateral medulla (CVLM)²³, the hypothalamus²⁴, the cortex²⁵ and the PAG-RVM complex²⁶.

The best-studied relay of descending modulation is the midline PAG-RVM complex²⁶, specific for noxious stimulation, which can both enhance and inhibit pain in different settings^{27,28}. As mentioned previously, the PAG receives inputs from limbic forebrain areas as the anterior cingulate cortex, AMY, hypothalamus, and mPFC 29 and relays this information to the spinal cord, through the rostral ventromedial medulla (RVM)³⁰. The RVM is a region of the ventral medulla that includes the midline nuclei raphe magnus (RMg), gigantocellularis reticularis (Gi), gigantocellularis pars alpha (GiA) and the paragigantocellularis lateralis

(LPGi). Projections from the RVM represent a major portion of the neurons projecting to the SDH. Electrophysiologically, the RVM is comprised of three different types of neurons, depending on their response to noxious stimulation, namely the pronociceptive On-, the antinociceptive Off- and the Neutralcells³¹.

When a noxious stimulus is applied, the activity of On-cells increases immediately before a withdrawal response while the opposite is observed for Off-cells. The third type of cells, Neutral-cells, do not alter their activity during peripheral noxious stimulation. Off-cells were first described to exert an antinociceptive action, as their activation is sufficient to produce analgesia³²⁻³⁴, as well as required for the action of opioids35,36. On the opposite side, on-cells are considered pronociceptive, since their activation induces hyperalgesia^{37,38}. The balance between the activation of on- and off-cell populations, therefore, allows a graded dual control of nociception²⁶.

Descending modulation occurs through three main processes: (i) a tonic effect, in which cutaneous sensory processing is inhibited, whereas pronociceptive activity is facilitated³⁹; (ii) an inhibitory effect, wherein supraspinal structures inhibit the activity dorsal horn neurons, acting indirectly through interneurons⁴⁰⁻⁴² or directly by inhibiting primary afferents^{43,44} and (iii) a facilitatory effect, where descending pathways enhance spinal nociceptive transmission from the SDH^{45,46}.

1.5. Chronic pain

Chronic pain results from an abnormal function of the nervous system, in which pain persists beyond healing time (more than 3-6 months)⁴⁷, and affects approximately 20% of the European population^{48,49}. This abnormal neuronal activity includes the sensitization of the peripheral and the central nervous systems⁵⁰, leading to a heightened perception of pain⁵¹. According to its aetiology, chronic pain can be divided into 7 categories⁴⁷: (i) primary pain, (ii) cancer pain, (iii) posttraumatic and postsurgical pain, (iv) neuropathic pain, (v) headache and orofacial pain, (vi) visceral pain and (vii) musculoskeletal pain (Table 1).

Table 1. Classification and definition of the most common clinically relevant chronic pain disorders, according to the 11th revision of the International Statistical Classification of Diseases and Related Health Problems (ICD) $^\pi$.

5

1.5.1. Neuropathic pain

As shown in Table 1, neuropathic pain is defined as *pain caused by a lesion or disease of the* somatosensory system² and is accompanied by the development of spontaneous pain, hyperalgesia (increased pain perception in response to a noxious stimulus) and allodynia (pain sensation in response to normally innocuous stimulus)⁵³. Although the exact prevalence of chronic neuropathic pain is unknown, it is estimated that it affects 7-10% of the population^{54,55}. Importantly, neuropathic pain greatly reduces the quality of life of patients, as it impairs physical, mental, emotional and social functioning⁵⁶. Regarding treatment, many of the patients are refractory to the currently available therapies, namely nonsteroidal anti-inflammatory drugs, opioids, and antidepressants^{57,58}. This leads to poorer quality of life for the patients⁵⁹, as well as higher use of healthcare resources and greater costs⁶⁰.

1.5.2. Emotional and cognitive impairments in neuropathic pain

Chronic pain goes beyond the sensory perception of pain, with sleep disturbances and emotional disorders, such as anxiety and depression, being frequently reported by patients⁶¹. Especially in neuropathic patients, psychiatric comorbidities have an estimated prevalence of 30%62. However, the mechanisms that underly the comorbidity between emotional disorders and neuropathic pain remain unclear.

Numerous studies showed negative emotions increase pain perception⁶³. Pain-related anxiety can increase pain sensitivity64–66 and worsen the subjective experience of pain67, predisposing people to develop a chronic pain condition68–70. Pain modulation by anxiety results from interactions between areas involved in both emotional and sensory processing such as the activity of the thalamus^{71,72}, amygdala^{72,73} and the PAG⁷³. Regarding major depressive disorders, epidemiological studies show a prevalence of approximately 50% in chronic pain patients^{74,75}. This comorbidity was associated with amplified pain perception⁷⁶, as well as

an increase in the activity of the amygdala, insula, and mPFC $77,78$ – areas shown to be involved in pain modulation.

In addition to emotional impairments, cognition also affects the experience of pain⁷⁹. Individuals with cognitive impairments display altered pain processing⁸⁰, however, pain sensitivity is variable according to the cause, which can be a consequence of ageing, neurological diseases or trauma⁸¹. Concerning attention, focusing on pain increases its perception decreasing the activity of areas including the thalamus and anterior cingulate cortex, and increasing the activity of others such as the PAG^{ω} – while a distraction can diminish it⁸³. Also, a verbal cue suggesting pain relief or increase can also decrease/increase pain experience^{84–86}, importantly this expectancy was also shown to alter analgesic efficacy^{87,88}. Fear of pain can cause persistent physical inactivity, leading to a negative spiral of worsened physical condition, frailty and distress, and subsequently more pain⁵⁹.

1.6. Animal models of neuropathic pain

The complexity of chronic pain, and especially of chronic neuropathic pain, has led to the development of a plethora of animal models, that can be categorized according to the location and method of injury **(Table 2)** into 5 gross categories^{89,90}: (i) central pain models, (ii) peripheral nerve injury models, (iii) models of disease-induced, (iv) drug-induced, and (v) inherited neuropathies. In rodents, the most common experimental approach is traumatic nerve injury (full or partial) via ligation, transection, or compression with the chronic sciatic constriction injury (CCI)⁹¹, spared nerve injury (SNI)⁹² and spinal nerve ligation (SNL)⁹³ models being mostly used.

Table 2. Animal models of neuropathic pain.

1.6.1. Chronic constriction injury model (CCI)

The CCI model mimics the symptoms of chronic nerve compression, such as nerve entrapment or compression syndromes158,159, comprising both the inflammatory¹⁶⁰ and neuropathic components161. This model, established in 1988 by Bennet and Xie91, results in hyperalgesia to thermal and mechanical stimuli with animals also displaying signs of spontaneous pain such as limping, limb guarding, excessive licking and avoidance of weight bearing on the injured limb^{91,161,162}. It is a reliable and reproducible model, that leads to the formation of intraneural oedema, axotomizing many but not all of the sciatic nerve axons163.

1.7. Emotional and cognitive impairments in the CCI model of neuropathic pain

In addition to the study of changes in nociception, animal models have also been used to study the anxiodepressive and cognitive components. Available behavioural evidence on the CCI model is summarized in Tables 3-4. Taking together, these studies show the development of anxiety- and depressive-like behaviours in the CCI model of neuropathic pain. Disregarding the general inference, results vary between studies (even between those using the same behavioural tests) concerning the time post-CCI at which they are observed and differences between tests performed on the same animals. This variability is mostly attributable to the use of different methodological approaches, namely the animal's species and strains, the behavioural paradigms and experimental designs used, which difficult the comparison between studies. Likewise, it becomes imperative the use of female animals as experimental subjects, as these studies mainly focus on male subjects and these comorbidities develop in a sex-specific manner, with special impact in women.

1.7.1. Anxiety-like behaviour

Evaluation of anxiety-like behaviour (Table 3) used mostly exploratory-based tests, which play on the inner struggle of the animals between approaching or avoiding novel situations and/or stimuli¹⁶⁴.

In the open field (OF) test, animals are placed in a square arena with an anxiogenic illuminated centre. Overall, CCI animals displayed a reduction in the time spent in the centre of the OF arena and in the number of entries in the central area, an indicator of anxiety-like behaviour¹⁶⁵⁻¹⁶⁹. Some exceptions¹⁶⁹⁻¹⁷¹ saw no differences in time spent in the centre of the arena of the OF. Identical results were obtained in the elevated plus maze (EPM), consisting of two opposing open and brightly lit arms and two opposing closed arms, in a "plus sign" shape, elevated above floor level. Anxiety is derived from a reduction in the number of entries and/or the time spent in the open arms, which was observed in the majority of the studies^{168,172–177}. In some exceptions^{168,171} no differences were found between control and CCI animals. One modified version of the EPM is the elevated zero maze178, which excludes the ambiguous central area in the traditional design179. Again, results were similar with CCI animals spending less time in the open quadrants, with one exception¹⁸⁰ which showed no differences between CCI and SHAM animals. Only one article used the light/dark box test (LDB)¹⁸¹, which uses a similar exploration/avoidance conflict as the OF, by allowing the animals to freely move between a dark and a bright chamber. A decrease in time spent in the lit compartment is an indicator of an anxiety state and, accordingly, CCI spent time in the bright compartment¹⁸².

The marble burying test (MB)¹⁸³ is based on the inherent burying behaviour of rodents on anxiogenic circumstances183, and an increase in the numbers of marbles buried is then considered as an indicator of anxious behaviour. In the two studies that used this paradigm, anxiety-like behaviour was not observed: Wilkerson et al.¹⁸⁴ showed a decreased in the number of marbles buried by CCI animals, whereas Bravo and colleagues¹⁸⁰ showed no differences.

The place escape/avoidance paradigm (PEAP) was used to evaluate an aversion to painful experiences¹⁸⁵: the animals are allowed to explore between a dark non-anxiogenic chamber, in which the injured paw was stimulated with a noxious stimulus, and a mildly bright anxiogenic side in which there is no stimulation or of the injured paw. In all studies, CCI animals spent less time in the dark chamber, showing an aversion to painful stimuli.

1.7.2. Depressive-like behaviour

For the study of depressive-like behaviours (Table 4), the forced swimming (FST), tail suspension (TST), and sucrose preference (SPT) tests are the most frequently used. The FST and TST (applied only in mice) evaluate the learned helplessness component of depression. In the FST186, the animals are placed in cylinders filled with water, in a no-escape situation. Similarly, in the TST¹⁸⁷ mice are suspended by the tail and similar struggling/immobility behaviours are assessed. In both tests, an increase in immobility behaviour is an indicator of behavioural despair. CCI animals displayed an increased immobility time but no differences in the time spent swimming, thus indicating the increase in immobility occurs at the expense of the decrease in the time spent climbing. The distinction between active behaviours (swimming/climbing) is important, as it is known to be differentially due to serotonergic and noradrenergic mechanisms, respectively188,189. Two exceptions were found190,191, in which no differences were seen in immobility, swimming and climbing time in the FST. Similarly, in the TST, an increase in immobility was also observed.

In the sucrose preference test (SPT), the animals are allowed to choose between a sweet (sucrose) and water bottle¹⁹². A decrease in the intake of sucrose indicates the development of anhedonic-like behaviour. This was observed on CCI animals^{165,193-196}, with one exception, where saccharin was chosen and no differences were found between sham and CCI animals on its preference¹⁶⁸.

In the novelty-suppressed feeding test (NFST), food-deprived animals are put in a chamber with a single pellet of food and the latency to approach it is recorded¹⁹⁷. This induces a conflict between the drive to eat and the fear of a novel environment. A hesitation to eat in a novel environment is described as intermix measure of anxiety and depression behavioural symptoms, observed in CCI animals^{182,193}.

Table 3. Summary of the literature concerning the development of anxiety-like behaviours in the chronic constriction injury model of neuropathic pain. Abbreviatures: OVX ovariectomized; M – male; F – female; L – left; R – right; ALB - anxiety-like behaviour; OF - open field test; EPM - elevated plus-maze; EZM - elevated zero-maze; LDB - light/dark box; MB - marble burying test; PEAT - place escape/avoidance test; SI - Social interaction; N/D – unknown.

Table 4. Summary of the literature concerning the development of depressive-like behaviours in the chronic constriction injury model of neuropathic pain. Abbreviatures: M male; F - female; L - left; R - right; DLB - Depressive-like behaviour; FST - forced swimming test; TST - tail suspension test; SPT - sucrose preference test; NSFT - novelty suppressed feeding test

1.8. Sex differences in pain perception and emotion

For the purpose of this thesis, the term sex will be used as it refers to biological sex, as gender is an identified sex that can be based on social or cultural values. Men and women do not experience pain in an equal manner. In chronic pain, women represent the majority of chronic pain patients, especially in chronic neuropathic pain^{55,216}, and have stronger and longer lasting pain symptoms²¹⁷. Differences have been also reported in the response to treatment, mainly concerning opioid analgesia, with women having a greater antinociceptive effect^{218,219}. In parallel, the prevalence of anxiety disorders and depression is also higher in women than in men and increases in the peri- and post-menopause period^{60,220}. Importantly, the severity of emotional disorders is also enhanced in women²²⁰.

In preclinical research, even though males are usually the subject of choice, sex must be taken into consideration, as previous studies showed neuropathic pain behaviours differ between males and females²²¹⁻²²³. In the CCI model of neuropathic pain, sex-related differences were observed in terms of nociception, with female rats experiencing higher thermal hyperalgesic symptoms as well as mechanical allodynia in comparison with males^{221,224,225}. Also, in mice, differences were observed in the development and recovery of neuropathic pain, as female mice showed higher mechanical allodynia in the first weeks post-CCI and males recovering faster from nerve injury²²⁴.

Even though chronic pain and affective disorders are more prevalent in women, there is a male-orientated bias in the choice of the experimental subject in basic research²²⁶ and female behaviour is presumed to be comparable to male behaviour. Consequently, a systematic review of the literature highlighted that the CCI model was never extensively characterized on female rats, especially regarding the development of mood disorders concomitant to neuropathic pain. The main goal of this work is, therefore, to characterize CCI-induced nociception and emotional impairments in female rats, comparing it with males with CCI. We will further analyse sex-specific changes in the neuronal activity of the RVM, an area involved in nociceptive and emotional processing. Taking this into consideration, this work was divided into three main goals:

(i) Characterization of CCI-induced nociceptive behaviour, in what concerns the development of mechanical and cold allodynia.

(ii) Characterization of CCI-induced emotional impairments, namely the development of anxiety- and depressive-like behaviours.

(iii) Assessment of changes in neuronal activity in the RVM, area implicated in nociceptive processing, through the analysis of single cell extracellular electrophysiology.

3.1. Animals and ethical issues

Adult Wistar Han rats (Charles Rivers, Barcelona, Spain), weighing between 200-350 g were used in all experiments, in a total of 38 animals. The animals were housed in groups of 2 in clear plastic cages with solid floors, covered with sawdust as bedding. Food and water were available *ad libitium* and animals were kept under a 12h light/dark cycle (artificially illuminated from 8h-20h), in a climate-controlled room (temperature: 20-24 $^{\circ}$ C; relative humidity: 55 +/- 10%). All manipulations were carried out during the light period of animals, between 9:00 a.m. and 5:00 p.m. Each animal was considered a single unit within its experimental group. The experimental protocol followed the European Community Council Directive 86/609/ EEC and 2010/63/EU concerning the use of animals for scientific purposes and was approved by the Institutional Ethical Commission (SECVS 16/2018). This work followed the ethical guidelines of the International Association for the Study of Pain regarding the use of laboratory animals and all efforts were made to minimize animal suffering and to use only the number of animals necessary to produce reliable scientific data.

3.2. Experimental design

Randomization for experimental group allocation was performed using a computer-generated randomization sequence (RANDOM.ORG). The animals were distributed into five groups: males with CCI $|CC|_M$, n=7), gonadally intact females (SHAM/SHAM, n=8), ovariectomized females (SHAM/OVX, n=7), gonadally intact females with CCI (CCI/SHAM, n=8) and ovariectomized females with CCI (CCI/OVX, $n=8$). The experimental timeline is illustrated in Figure 3. Before starting the experiments, the animals were handled daily for a minimum of 2 weeks to habituate to the experimenter. Ovariectomies were performed 3 weeks before CCI surgery. The week before CCI, a baseline evaluation of nociceptive behaviour was performed using the von Frey and acetone tests. After CCI surgery, nociception was evaluated weekly, until the end of the behavioural characterization. On week 5 post-CCI surgery, the animals were tested in the OF, EPM, SPT and FST tests. Histological and electrophysiological studies started at the end of the behavioural characterization. These experiments were performed through three independent sets to adjust the time period post-CCI surgery required for the electrophysiological recordings.

Figure 3. Schematic representation of the experimental design. Before the beginning of the experiment, all animals were handled daily by the experimenter for two weeks and, on the day of the experimental sessions, animals were left in the experimental room for an hour in order to habituate to the surroundings. After habituation, female animals underwent ovariectomy surgery and, three weeks later, chronic constriction injury induction or sham surgery, accordingly to the correspondent group. Identically, male animals underwent chronic constriction injury induction. Nociceptive behaviour was evaluated weekly, starting one week before induction. After four weeks, emotional-like behaviours were assessed, namely anxiety- and depressive-like behaviour. Following behavioural evaluations, the animals underwent electrophysiological recordings or were sacrificed, and samples were processed for histology. (CCI – chronic constriction injury; OF – open field test; EPM – elevated plus-maze; SPT – sucrose preference test; FST – forced swimming test; W - week).

3.3. Anaesthesia protocol

For both CCI and ovariectomy surgeries, the animals were anaesthetized intraperitoneally (i.p.) with a mixture of ketamine (0.75 mg/kg, Imalgene, Merial, Oeiras, Portugal) and medetomidine (0.5 mg/kg Dorbene, Esteve, Carnaxide, Portugal). At the end of the surgery, the anaesthesia was reversed with atipamezole hydrochloride (1 mg/kg, Antisedan, Pfizer, Oeiras, Portugal, i.p.). Vaseline was applied to both eyes to protect them from drying during surgery and recovery. After both surgeries, the animals were monitored until fully awake (eating and grooming) and recovered, and welfare was monitored daily (grooming, wound appearance, dehydration/weight loss and locomotor abnormalities).

During electrophysiological recordings, anaesthesia was induced (50 mg/kg, Eutasil, Ceva, Algés, Portugal) and maintained through intraperitoneal injection of pentobarbitone. The level of anaesthesia was monitored by verifying the dilation of the pupils, the general muscle tone and responses to noxious pinching every 15 min. An additional dose of pentobarbitone (20 mg/kg) was given if observed a pinchevoked response.

3.4. Ovariectomy

Females were ovariectomized (OVX) bilaterally²²⁷. After a single mid-dorsal skin incision at the level of L2-L6, the cutaneous maximus and external oblique, internal oblique and transversus abdominis muscles were transected, 1 to 2 cm in length, about 1 cm caudal to the last rib. Each ovary was exteriorized through the incision and the oviducts and ovarian vessels were ligated using 4-0 silk sutures (Mersilk, Ethicon, New Jersey, EUA) and the entire ovary removed. Surgical incisions were closed in layers and the anaesthesia reverted. The same procedure was carried out for the SHAM animals except for the removal of the ovaries.

3.5. Induction of neuropathic pain

The chronic constriction injury (CCI) model was induced as originally described⁹¹. Briefly, the common left sciatic nerve was exposed by blunt dissection of the biceps femoris. Proximal to the sciatic nerve's trifurcation, four 3/0 chromic gut ligatures (B. Braun Surgical, Rubi, Spain) were tied loosely with approximately 1 mm spacing, placed as to not arrest circulation. Sham animals received all the surgical procedures, but the sciatic nerve was left without ligation. Afterwards, surgical incisions were closed in layers and the anaesthesia reverted.

3.6. Behavioural assessment of nociception

3.6.1. Mechanical allodynia

The von Frey test was used for mechanical allodynia assessment, using a series of calibrated von Frey monofilaments (North Coast Medical Inc, Morgan Hill, CA). The animals were placed in transparent plastic containers on an elevated mesh floor and allowed to adjust to the environment. Von Frey filaments were applied to the plantar surface of each hindpaw in a series of ascending forces (equivalent to 0.4, 0.6, 1, 2, 4, 6, 8 and 15 g force), initiating with the 2 g filament. Positive responses included prolonged paw withdrawal, followed by licking or scratching. The 50% response threshold was calculated using the updown method²²⁸.

3.6.2. Cold allodynia

Cold allodynia was evaluated through the acetone test. In the same apparatus as the von Frey test, a 100 µL drop of acetone was applied with a pipette in the centre of the ventral surface of each hindpaw. The response was graded in a three-point scale: 0, no response; 1, quick withdrawal, flick or stamp of the paw; 2, prolonged withdrawal or repeated flicking of the paw; 3, repeated flicking of the paw with persistent licking directed at the ventral side of the paw. Acetone was applied alternately 3 times, at 1 min intervals, and the mean score value was calculated.

3.7. Behavioural assessment of emotional behaviour

3.7.1. Anxiety-like behaviour

3.7.1.1. Open field test (OF)

The OF test was used to evaluate anxiety-like behaviour, following a previously described protocol²²⁹. Briefly, the test is performed in a square arena (Med Associates Inc., St. Albans, Vermont, USA), 43.2 cm wide, with transparent acrylic walls and brightly illuminated in the centre (240 lx). The animal is placed in the centre of the arena (corresponding to a square 21.6 cm wide, equidistant from the borders) and its activity is automatically recorded for 5 min using the Activity Monitor 5 software (Med Associates Inc., St. Albans, Vermont, USA). The locomotor capability was assessed using the total distance travelled by the animals as well as average velocity, and anxiety-like behaviour through the percentage of time spent in the centre of the arena. Between each trial/animal, the arena was cleaned thoroughly with 10% alcohol.

3.7.1.2. Elevated plus maze (EPM)

Anxiety-like behaviour was also evaluated through the EPM192. The apparatus is composed of two open arms (50.8 cm \times 10.2 cm) and two opposing closed arms (50.8 cm \times 10.2 cm \times 40.6 cm), elevated 72.4 cm above the ground (ENV-560; Med Associates, St. Albans, Vermont, USA). The animals were placed in the centre of the maze and allowed to explore for 5 min. Behaviour was recorded and then analysed using the EthoVision XT 13 software (Noldus Information Technology, Wageningen, Netherlands). Anxiety-like behaviour was assessed based on the time spent in the open arms, expressed as a percentage of the total testing time. The total number of arm entries was also evaluated. The maze was cleaned with 10% alcohol between trials/animals.

3.7.2. Depressive-like behaviour

3.7.2.1. Anhedonic-like behaviour

To evaluate anhedonic-like behaviour, the sucrose preference test (SPT) was performed¹⁹², with minor modifications. One week before surgery, the animals were pre-exposed to the sucrose solution. During their active period, the animals were given free access to both water and a sucrose solution (2%) for 2h, and efforts were made to ensure that all animals tasted the solution. On test day, the animals were presented with two pre-weighted bottles (water and sucrose) for 3h. Afterwards, the bottles were weighted, and the sucrose preference calculated as follows:

Successe Preference
$$
(\%) = \frac{\text{success intake}}{(\text{success + water}) \text{ intake}} \times 100
$$

To control for potential bias associated with differences in body weight between experimental groups, sucrose preference was additionally adjusted to the animal's body weight. The total intake of sucrose and water was also assessed and likewise adjusted to the animal's body weight on testing day.

3.7.2.2. Learned helplessness

Learned helplessness was evaluated using the Forced Swimming Test (FST)¹⁹². The day before testing, the animals were submitted to a pre-test session, in which they were placed individually in a cylinder (diameter: 29 cm; height: 50 cm) filled with water (approximately 25ºC, 30 cm in depth). After 10 min, the rats were removed from the water and towel-dried before returning to their home cages. On testing day, animals were again placed in the cylinders, for a period of 5 min, and the session recorded. Using the freeware Etholog software²³⁰, latency to immobility, time spent immobile, swimming and climbing were quantified. Learned helplessness behaviour was defined by an increase in the time of immobility at the expense of the time spent swimming/climbing.

3.8. Electrophysiological recordings in the RVM

For the electrophysiological study, the animals were removed from the animal house randomly, one each day, and brought to the recording room. The depth of anaesthesia was assessed using the tail-pinch reflex, and recordings started when the animal was under light anaesthesia, neurones responded to noxious stimulation but no behavioural response to noxious pinching of the tail was observed. Body temperature was maintained with the help of a warming blanket (DC Temperature Controller, FHC, Bowdoin, ME, USA).

For supraspinal recordings in the RVM, the animals were initially placed in a standard stereotaxic apparatus. The skull was then exposed and an osteotomy was performed, to allow the placement of the recording electrodes according to the atlas of Paxinos and Watson²³¹ (AP: 10.92 mm caudal to the bregma; ML: 0.00 mm from the midline; DV: 10.0 mm below the surface of the skull). Single neuron activity was recorded extracellularly with tungsten electrodes (tip impedance 3–10 MV at 1 kHz), the signal was amplified and filtered, and data sampling was performed through a CED Micro 1401 interface and Spike 2 software (Cambridge Electronic Design, Cambridge, UK).

The response of RVM cells to peripheral stimulation (applied to the left hindpaw) was evaluated through the following assessments, performed successively: (i) spontaneous activity; (ii) response to brushing; (iii) response to mechanical stimulation using von Frey hair, applied once (6 gf). (iv) response to pinching of the tail with a surgical clamp; (v) response to cold stimulation of the plantar skin. After finishing a recording, the animals were allowed a resting period of about 30 min starting a new recording.

To calculate the evoked response (∆ Activity) of the recorded neurons, the baseline discharge frequency was subtracted from the evoked activity using the following equation:

 \triangle Activity = (cell activity during stimulation) – (basal cell activity prior to stimulation)

Then, RVM neurons were classified as WDR-like or NS-like according to their response to innocuous mechanical stimulation (brushing), being responsive or non-responsive, respectively. Neurons were then classified for each stimulus as (i) On-like if their activity increased during stimulation, (ii) Off-like cells if their activity decreased during the stimulation period and (iii) Neutral if none or negligible changes (<10%) were observed. This classification varied from that of Fields and colleagues³¹, as the noxious-induced withdrawal reflex was not taken into consideration and only changes in neuronal activity were considered. Therefore, neurons are herein referred to as On-like and Off-like cells rather than On- or Off- cells^{232,233} (Figure 4).

Figure 4. Example of an output from the recording software Spike 2. (A) Example of a recording of an NS On-like neuron activity. While not responding to innocuous stimulation (green interval), the cell increases its firing rate during the noxious stimulus (orange interval). (B) Sample recording of a WDR Off-like neuron. The activity of this cell is decreased not only by an innocuous stimulus (green interval) but also by noxious stimulation (orange interval). (C) Example of a recording from a neutral cell. This cell does not respond to either innocuous or noxious stimulus (Green lines – innocuous mechanical stimulation; Red lines – noxious mechanical stimulation to the tail).

After completing a recording session of approximately 4-6 hours, the animals were given a lethal dose of pentobarbitone (80 mg/kg, i.p.; Eutasil, CEVA) and were perfused with 4% paraformaldehyde (PFA).

3.9. Histology

3.9.1. Vaginal Smears

Female animals were monitored for their oestrous cycle using vaginal smears collected during habituation and on each behavioural day, after testing. In brief, for sample collection, a drop of saline is placed in a pre-labelled glass slide. A smear loop is moistened in saline and the female rat is held around the thorax, lightly gripping the tail with the same hand. The tip of the loop is then inserted carefully, with a small rotation movement, and rolled gently in the glass slide. Lastly, the animal is allowed to calm down and rest in its home cage. The samples were air-dried and fixed with 95% ethanol, followed by a Papanicolaou stain. Slides were evaluated using a brightfield microscope (Olympus Widefield Upright Microscope BX61) at the original objective magnifications of 4X, 10X and 20X (Figure 5).

Figure 5. Sample images of the vaginal smears analysed. Represented are the (A) proestrus, (B) oestrus, (C) metestrus and (D) dioestrus phases of the oestrous cycle in female Wistar Han rats, stained with the Papanicolaou stain. Original magnification of 10X (scale: 100 µm).
The oestrous cycle was classified between 4 stages: proestrus, oestrus, metestrus and dioestrus based on the presence or absence of nucleated epithelial cells, cornified epithelial cells and leukocytes234. In the proestrus stage, there is a predominance of nucleated epithelial cells, while oestrous is distinctively characterized by cornified squamous epithelial cells. In metestrus, there is a mix of leucocytes and cornified squamous epithelial cells, whereas the dioestrus stage consists predominantly of leukocytes.

3.9.2. H&E staining of the sciatic nerve

Sections of the sciatic nerve were embedded in a paraffin block and sectioned into 5 μm-thick slices with a microtome. After staining with haematoxylin and eosin (H&E), slides were examined using a light microscope (Olympus Widefield Upright Microscope BX61) regarding abundance, size, and arrangement of nerve fibres.

3.10. Statistical analysis

The GraphPad Prism 6 software (GraphPad Software Inc, La Jolla, CA, USA) was used to perform the statistical analysis and graphical representations. Body weight changes and nociceptive data were analysed using a two-way repeated measures analysis of variance (ANOVA), comparing all the experimental groups. Data analysis of emotional, electrophysiological and histological data was performed using a two-way ANOVA followed by a Bonferroni test for post-hoc multiple comparisons comparing the four female groups, with ovariectomy and CCI as variables. A t-test for unpaired data with Welch's correction was used to compare CCI males with CCI females (CCI/SHAM and CCI/OVX females). Results were expressed as the mean ± standard error of the mean (SEM). In all cases, P<0.05 was considered statistically significant.

23

4.1. Animal Welfare

CCI animals walked with a definite limp, often guarding the affected hindpaw. While usually the heel is elevated, CCI animals stood with the hindpaw everted and the heel touching the floor. The toes, which are normally spread apart, were together and ventroflexed while walking or standing. Occasionally, autotomy was observed, with gnawed claw tips. These behaviours were not observed SHAM/SHAM and SHAM/OVX animals.

Weight change after CCI surgery is represented in Figure 6. This parameter was considered a measure of the general welfare of the animals, and no precipitous weight loss or dehydration was observed. This value was calculated in relation to the animals' body weight the week before surgery (baseline).

Figure 6. Variation of body weight throughout the experiments, starting one week before CCI surgery. CCI males, as well as SHAM/SHAM and SHAM/OVX females, showed an increase in body weight, while CCI females showed an arrest in weight gain. Results are presented as mean \pm SEM. *P<0.05 and ***P<0.001 vs. baseline (week 0) within the respective group (t-test with a Bonferroni correction for multiple comparison t-test with a Bonferroni correction for multiple comparisons). Symbols are coloured according to their respective experimental group. $(n_{cc/M} = 7, n_{shAM/SHAM} = 8, n_{shAM/OVX} = 7, n_{cc1/SHAM} = 8$ and $n_{cc1/OVX} = 8$.

Statistical analysis showed body weight varied differently throughout time, depending on the experimental group (Interaction: $F_{21,182} = 6.34$, P<0.001, $\eta_{\rho}^2 = 0.12$; Time: $F_{7,182} = 23.3$, P<0.001, $\eta_{\rho}^2 = 0.15$; Group:

 $F_{3,26}$ =7.68, P<0.001, η_{p} and 27). Post-hoc tests showed that, while males, SHAM/SHAM and SHAM/OVX females increased their body weight throughout the behavioural period, CCI females displayed no significant weight gain during the evaluation period, with no differences between CCI/SHAM and CCI/OVX females (P>0.99). By contrast, SHAM/OVX showed a significant increase in body weight starting at week two post-CCI surgery (P<0.001).

4.2. Behavioural assessment of nociception

To evaluate the development of nociceptive-related behaviours, the von Frey test was performed to assess mechanical allodynia and the acetone test for cold allodynia. These tests began the week before CCI surgery (baseline) until week 5 after surgery, the endpoint for behavioural assessments (**Figure 7**).

Figure 7. Assessment of nociceptive behaviour throughout the experimental period. (A) Mechanical allodynia was observed from week 1 and 2 onwards in CCI/OVX and CCI/SHAM female rats, respectively, with significant differences between the two experimental groups on weeks 1 and 5. (B) Regarding cold allodynia, CCI females displayed cold allodynia from week 1 onwards, in comparison with SHAM animals. CCI males displayed both mechanical and cold allodynia from week 1 onwards but recovering at week 5. Results are presented as mean \pm SEM. *P<.05, **P<0.01 and ***P<0.001 vs. baseline (week 0) within the respective group (t-test with a Bonferroni correction for multiple comparison t-test with a Bonferroni correction for multiple comparisons). Symbols are coloured according to their respective experimental group. $(n_{c c l/M} = 7, n_{s HAM/S HAM} = 8, n_{s HAM/OV X} = 7, n_{c c l/S HAM} = 6-7$ and $n_{cc_{1}/\text{ovx}} = 7$).

4.2.1. Mechanical allodynia

Regarding mechanical allodynia, statistical analysis showed this parameter varied differently between experimental groups throughout time (Figure 7A; Interaction: F_{20,160}=7.23, P<0.001, η_ρ²⁼0.24; Time: $F_{5,160} = 11.5$, P<0.001, $\eta_{\rho}^2 = 0.094$; Group: $F_{4,32} = 39.3$, P<0.001, $\eta_{\rho}^2 = 0.34$). Post hoc tests showed a significant decrease in withdrawal thresholds in CCI females, starting at week 1 for CCI/OVX animals and

at week 3 post-CCI surgery for CCI/SHAM females. Interestingly, while CCI/OVX females developed mechanical allodynia earlier than CCI/SHAM (P=0.041), these females appear to recover on week 5 unlike CCI/SHAM females (P=0.035).

CCI males showed a decrease in the withdrawal threshold, an indicator of the development of mechanical allodynia, starting from week 1 onwards when compared with baseline values. After week 3, recovery started to be observed in neuropathic males and, at week 5, no significant differences were observed in comparison with baseline. In comparison with neuropathic females, males were shown to have lower withdrawal thresholds at weeks 1 (CCI/SHAM: P<0.001; CCI/OVX: P=0.003) and 2 (CCI/SHAM: P<0.001; CCI/OVX: P<0.001). Oppositely, at week 5 CCI_M showed higher withdrawal thresholds than CCI/SHAM females (P<0.001) but no differences in comparison with CCI/OVX (P>0.99).

4.2.2. Cold allodynia

In the acetone test, which assesses the development of cold allodynia, ANOVA analysis showed this parameter varied differently throughout time between experimental groups (Interaction: $F_{20,150}=7.55$, P<0.001, η_{ρ}^2 =0.15; Time: F_{3,150}=29.2, P<0.001, η_{ρ}^2 =0.14; Group: F_{4,30}=45.7, P<0.001, η_{ρ}^2 =0.49). Post hoc tests confirmed CCI_M, CCI/SHAM and CCI/OVX experimental groups cold allodynia from week 1 onwards (Figure 7B).

However, and contrary to what is observed for CCI females, neuropathic males steadily recovered after week 3 post-CCI induction. Cold allodynia in CCl_{M} was shown to be different from CCI/SHAM at weeks 1 (increased, P=0.044), 3 (decreased, P=0.045) and 5 (decreased P<0.001), as well as from CCI/OVX at week 5 (decreased, P<0.001).

4.3. Anxiety-like and locomotor behaviour

To assess the development of anxiety-like behaviour at week 5 after CCI surgery, the OF and EPM tests were performed. Basal locomotion was also evaluated through the OF test.

4.3.1. Open-field test (OF)

Anxiety-like behaviour was assessed in the OF by counting the number of *faecal boli* (Figure 8A) and assessing the time spent in the centre of the arena (Figure 8B). Two animals were excluded from the analysis due to software malfunction during the test (one from CCI/SHAM and CCI/OVX groups, respectively).

26

Figure 8. Evaluation of locomotion and anxiety-like behaviour in the open-field (OF) test four weeks after CCI surgery. (A) The number of faecal boli left in the arena at the end of the trial. (B) Time spent in the centre during the OF test expressed as a percentage (%) of total testing time. (C) Total distance (cm) travelled in the open field arena. (D) Average velocity during the trial. CCI/OVX animals showed a decrease in the total distance travelled, suggesting locomotor impairments Neuropathic males showed increased time spent in the centre and distance travelled in comparison with CCI/OVX females. Results are presented as mean \pm SEM. *P<0.05 (t-test with a Bonferroni correction for multiple comparisons). # $P<0.05$ (unpaired t-test with Welch's correction). $(n_{\text{c}}=7,$ $n_{\text{sham/sham}} = 8$, $n_{\text{sham/ovx}} = 7$, $n_{\text{cc1/sham}} = 7$ and $n_{\text{cc1/OVx}} = 7$).

Regarding the number of *faecal boli* left in the arena, CCI significantly increased anxiety-like behaviour in females (F_{1,25}=4.75, P=0.039, η_β²=0.16), independently of ovariectomy (F_{1,25}=0.58, P=0.46, η_β²=0.019) however post hoc analysis showed no differences between female experimental groups. Comparison between males and neuropathic females also showed no significant differences.

Neither CCI nor ovariectomy altered the time spent in the centre of the arena (CCI: $F_{1,25}=0.066$, P=0.80, η_{ρ}^2 =0.023; OVX: F_{1,25}=2.57, P=0.12, η_{ρ}^2 =0.092). Interestingly, CCI_M spent a higher percentage of time in the centre of the OF (t_7 =2.53, P=0.039), in comparison with CCI/OVX females although no differences were found in relation to CCI/SHAM animals $(t_{11}=0.96, P=0.034)$.

Basal locomotion was assessed in the OF test by analysing the distance travelled and average velocity of the animals. Regarding the distance travelled (**Figure 8C**), both ovariectomy and CCI independently decreased this parameter (CCI: $F_{1,25} = 6.83$, P=0.015, $\eta_{p}^2 = 0.19$; OVX: $F_{1,25} = 4.55$, P=0.043, $\eta_{p}^2 = 0.12$;

Interaction: $F_{1,25}$ =0.076, P=0.78, η_{p} ²=0.002). Further comparisons showed CCI/OVX animals displayed a significant decrease in the distance travelled, compared with the SHAM/SHAM group as well as in comparison with CCI_M (t₁₁=2.76, P=0.018). Nevertheless, no differences were seen between CCI_M and CCI/SHAM females $(t_{11}=0.96, P=0.36)$.

Regarding the average velocity (Figure 8D), this parameter was only decreased when animals were both OVX and CCI (Interaction: F_{1,25}=4.253, P=0.049, η_β²=0.13; CCI: F_{1,25}=0.29, P=0.59, η_β²=0.009; OVX: $F_{1,25}$ =2.68, P=0.11, η_{p} ²=0.084), and no significant differences were found between experimental groups.

4.3.2. Elevated plus maze (EPM)

The EPM was also used to evaluate anxiety-like behaviour through the *faecal boli* left at the end of the trial, as well as the time spent and entries in the open arms ([Figure 9](#page-41-0)). One CCI/SHAM animal was excluded from analysis after falling from the EPM apparatus.

Figure 9. Behavioural evaluation of anxiety-like behaviour in the elevated plus maze (EPM) test four weeks after CCI surgery. (A) The number of *faecal boli* left in the arena at the end of the trial. (B) Time spent in the open arms during the EPM test expressed as a percentage (%) of total testing time. (C) Total of open arms entries during the trial. CCI_M left fewer faecal boli and spent less time in the open arms that CCI/OVX and CCI/SHAM females. Results are represented as mean \pm SEM. # P<0.05 (unpaired t-test with Welch's correction). $(n_{\text{cc}}=7, n_{\text{SHAM} \sim \text{HAM}}=8,$ $n_{\text{sham/ovx}}$ =7, $n_{\text{ccl/sham}}$ =7 and $n_{\text{ccl/ovx}}$ =8)

Neither ovariectomy nor CCI induction altered the performance of females in the EPM regarding the number of *faecal boli* (Figure 9A; Interaction: F_{1,26}=0.79, P=0.38, η_β2=0.003; CCI: F_{1,26}=0.49, P=0.49, η_ρ = 0.018; OVX: F_{1,26}=0.002, P=0.96, η_ρ = 0.008), number of open arm entries (**Figure 9B;** Interaction: $F_{1,26}=0.0003$, P=0.99, η_{p} ²<0.001; CCI: $F_{1,26}=0.14$, P=0.71, η_{p} ²=0.005; OVX: $F_{1,26}=1.55$, P=0.22, η_{p} ²=0.056) and time spent in open arms (Figure 9C; Interaction: $F_{1,26}$ =0.003, P=0.96, η_{ρ} <0.001; CCI: $F_{1,26}$ =0.29, P=0.60, η_{p}^2 =0.011; OVX: $F_{1,26}$ =0.18, P=0.59, η_{p}^2 =0.007).

When comparing CCI_M with neuropathic females, males showed a decrease in the number of faecal boli in comparison with CCI/OVX females (t₇=2.39, P=0.047) but not with CCI/SHAM (t₆=1.72, P=0.14), as well as in time spent in the open arms in comparison with both CCI/SHAM (t =2.69, P=0.031) and CCI/OVX neuropathic females $(t_s=2.90, P=0.020)$.

4.4. Depressive-like behaviour

In order to assess depressive-like behaviour, on week 5 post-CCI surgery the SPT was performed as a measure of anhedonia and the FST to evaluate learned helplessness.

4.4.1. Sucrose Preference Test (SPT)

Sucrose preference was not altered neither by CCI nor ovariectomy (Figure 10A; Interaction: $F_{1,27}=3.42$, P=0.075, η_β²⁼0.10; CCI: F_{1,27}=0.011, P=0.92, η_β^{2<}0.001; OVX: F_{1,27}=3.99, P=0.056, η_β²⁼0.12).

However, when adjusted for the body weight of the animals (Figure 10B), ovariectomy independently decreased sucrose preference (OVX: $F_{1,27}=28.4$, P<0.001, $\eta_{\rho}^2=0.49$), indicating the development of depressive-like behaviour in ovariectomized females, while CCI did not induce anhedonic-like behaviour (CCI: $F_{1,27} = 0.024$, P=0.88, η_{p} <0.001; Interaction: $F_{1,27} = 3.02$, P=0.094, η_{p} ²=0.052). Confirming these results, multiple comparisons showed both SHAM/OVX and CCI/OVX animals decreased sucrose preference, in comparison with SHAM/SHAM animals. Males with CCI also displayed lower sucrose preference in comparison with CCI/SHAM $(t_{12}=7.47, P<0.001)$ and CCI/OVX females $(t_{12}=2.33 P=0.038)$.

To assess if there were differences in drinking patterns, the total intake of sucrose and water was evaluated. ANOVA analysis showed an interaction between CCI and ovariectomy (Figure 10C; Interaction: F_{1,27}=10.3, P=0.003; $\eta_{p}^{2}=0.23$; OVX: F_{1,27}=4.98, P=0.034; $\eta_{p}^{2}=0.11$; CCI: F_{1,27}=2.72, P=0.11, η_{ρ} ^{2=0.060). Multiple comparisons showed, in comparison with SHAM/SHAM animals, a decrease in the} total intake of SHAM/OVX and CCI/SHAM animals. When the intake was adjusted to body weight (Figure **10D**), identical results were observed in the ANOVA analysis (Interaction: $F_{1,27}=10.3$, P=0.003, $\eta_{\rho}^2=0.20$; OVX: F_{1,27}=11.2, P=0.002, η_β²=0.23; CCI: F_{1,27}=3.79, P=0.06, η_β²=0.072). Multiple comparisons showed a decrease in total intake in all female experimental groups, in comparison with SHAM/SHAM animals. In both these analyses, no differences were found between CCI_{M} and neuropathic females.

Figure 10. Behavioural evaluation of depressive-like behaviour using the sucrose preference (SPT) test four weeks after CCI surgery. (A) Sucrose preference in the SPT and (B) adjusted to body weight. (C) Total intake of water and sucrose solution in the SPT test and (D) Total intake adjusted to body weight. Ovariectomized female rats, with (CCI/OVX) and without CCI (SHAM/OVX) showed a decrease in sucrose preference, indicative of depressivelike behaviour. Results are presented as mean \pm SEM. **P<0.01 and ***P<0.001 (t-test with a Bonferroni correction for multiple comparisons). # P<0.05 and ### P<0.001 (t-test for paired data with Welch's correction). $(n_{\text{cell}}=7, n_{\text{SHAM/SHAM}}=8, n_{\text{SHAM/OVX}}=7, n_{\text{CCL/SHAM}}=8$ and $n_{\text{CCL/OVX}}=8$).

4.4.2. Forced Swimming Test (FST)

The FST was used to assess learned helplessness-like behaviour, a component of depression (Figure [11](#page-44-0)). Three different parameters were evaluated, namely the number of *faecal boli*, latency to immobility and immobility time. Time spent swimming and climbing is also represented in Figure 11C.

ANOVA analysis showed neither CCI nor ovariectomy influenced the females performance regarding (i) the number of *faecal boli* (Figure 11A; Interaction: F_{1,27}=0.20, P=0.66, η_{ρ} ²=0.097; CCI: F_{1,27}=0.20,

P=0.66, η_{ρ}^2 =0.007; OVX: F_{1,27}=0.37 P=0.55, η_{ρ}^2 =0.013); (ii) latency to immobility (Figure 11B; Interaction: F_{1,27}<0.001, P=0.98, η_β < 0.001; CCI F_{1,27}=2.34, P=0.63, η_β = 0.009; OVX: F_{1,27}=0.010, P=0.92, η_{ρ} <0.001) and (iii) immobility time (Figure 11C; Interaction: F_{1,27}=0.63, P=0.43, η_{ρ} =0.022; CCI: $F_{1,27} = 0.11$, P=0.74, $\eta_{p}^2 = 0.004$; OVX: $F_{1,27} = 0.65$, P=0.43, $\eta_{p}^2 = 0.023$). Further comparisons between neuropathic males and females showed no significant differences for all the parameters evaluated.

Figure 11. Behavioural evaluation of depressive-like behaviour using the forced swimming (FST) test four weeks after CCI surgery. (A) Number of faecal boli left at the end of the trial. (B) Latency to immobility. (C) Time spent climbing, swimming and in immobility measured in the FST. No significant differences were observed between experimental groups. Results are represented as mean \pm SEM. ($n_{\text{cell}}=7$, $n_{\text{SHAM}>\approx 8}$, $n_{\text{SHAM}>\approx 7}$, $n_{\text{cell}>\approx 8}$ and n $_{cc\rightarrow\sigma\sigma}$ =8).

4.5. Electrophysiological recordings in the RVM

A total of 447 neurons were recorded in the RVM. As described in the methods section, neurons were initially classified as WDR-like or NS-like according to their response to innocuous mechanical stimulation, namely brushing of the animal's back (Table 5). For the sake of simplicity, these cells will from now on be designated as WDR and NS.

Table 5. Total number of RVM WDR- and NS-like neurons recorded in the electrophysiological study. (WDR - Widedynamic range; NS - nociceptive-specific; n – number of animals recorded).

The spontaneous activity of both WDR- and NS-like cells in the RVM is represented in [Figure 12.](#page-45-0) ANOVA analysis showed no effects on their spontaneous activity (Table 6), and no differences were observed between neuropathic CCI/SHAM females and CCI_M.

Figure 12. Spontaneous activity of WDR and NS cells recorded in the RVM. Results are presented as mean \pm SEM. (WDR - Wide-dynamic range; NS - nociceptive-specific).

Table 6. Results of the statistical analysis of the differences in the spontaneous activity of RVM WDR and NS cells amongst experimental groups (WDR - Wide-dynamic range; NS - nociceptive-specific; CCI - chronic constriction injury).

Neurons were further classified as On-, Off- or Neutral-like cells according to their response for each stimulus applied and only On- and Off-like cells were analysed in this work.

4.5.1. Mechanical stimulation: Von Frey

RVM neuronal activity was evaluated through Von Frey mechanical stimulation (6 g). Classification and the total number of neurons responsive to von Frey stimulation recorded are described in [Table 7](#page-46-0).

Table 7. Total number of RVM On, Off- and Neutral-like cells to mechanical stimulation with Von Frey filament.

Spontaneous activity of RVM neurons prior to von Frey stimulation is represented in [Figure 13](#page-46-1). ANOVA analysis showed no differences in the spontaneous activity of all cell types between females (Table 8). By contrast, the spontaneous activity of NS Off-like cells was increased in males in comparison with both $CCI/SHAM$ (t_s=3.25, P=0.020) and CCI/OVX females (t_s=3.68, P=0.012)

Figure 13. Spontaneous activity of RVM neurons before mechanical stimulation with von Frey hairs. No significant differences were observed between experimental groups. Results are presented as mean \pm SEM. # P<0.05 (unpaired t-test with Welch's correction). (WDR - Wide-dynamic range; NS - nociceptive-specific).

Table 8. Results of the statistical analysis of the differences in the spontaneous activity of RVM neurons before mechanical stimulation with von Frey hairs between female experimental groups. (WDR - Wide-dynamic range; NS - nociceptive-specific; CCI - chronic constriction injury).

	Interaction	Ovariectomy	CCI
WDR On-like cells	$F_{1.36}$ =0.001, P=0.87, η _s <0.001	$F_{1,36}$ =0.22, P=0.64, η_{μ} ² =0.005	$F_{1.36} = 1.10$, P=0.30, $\eta_{\text{s}} = 0.029$
WDR Off-like cells	$F_{1.35}$ =0.56, P=0.46, η _s ² =0.015	$F_{1.35}$ =0.59, P=0.45, η_{μ} ² =0.016	$F_{1.35}$ =0.11, P=0.74, η_s ² =0.003
NS On-like cells	$F_{1,29}$ =0.44 P=0.51, η_{p} ² =0.015	$F_{1,29}$ =0.42, P=0.52, η _s ² =0.014	$F_{12} = 0.002$, P=0.96, n ² <0.001
NS Off-like cells	$F_{1,22}$ =0.38, P=0.54, η_{ρ} ² =0.016	$F_{1.22}$ =0.009, P=0.92, η _s < 0.001	$F_{1,22}$ =1.72, P=0.20, η _s ² =0.071

During Von Frey stimulation ([Figure 14](#page-47-0)), ovariectomy independently increased the evoked response of WDR Off-like cells (Table 9). Multiple comparisons showed a significantly increased in WDR Off-like cells of CCI/OVX animals, in comparison with CCI/SHAM (P=0.015) and SHAM/SHAM (P=0.007) females, as well as in comparison with males $(t_{11}=3.07, P=0.011)$. No differences were observed for NS and WDR On-like neurons.

Figure 14. Evoked response of RVM neurons after mechanical stimulation with von Frey hairs. CCI/OVX showed an increased response of WDR Off-like cells in comparison with CCI/SHAM and SHAM/SHAM females. Results are presented as mean ± SEM. *P<0.05 and **P<0.01 (t-test with a Bonferroni correction for multiple comparisons). (WDR - Wide-dynamic range; NS - nociceptive-specific).

Table 9. Results of the statistical analysis of the differences in the evoked response of RVM neurons after mechanical stimulation with von Frey hairs between female experimental groups. *P<0.05. (WDR - Wide-dynamic range; NS - nociceptive-specific; CCI - chronic constriction injury).

4.5.2. Mechanical stimulation: Pinch

RVM neuronal activity was also assessed after noxious mechanical stimulation, namely through pinching of the tail using a surgical clamp. Classification and the total number of neurons recorded is described in

[Table 10](#page-48-0).

Table 10. Total number of RVM On-, On- and Neutral-like cells to noxious mechanical stimulation to the tail (pinching with a surgical clamp), recorded in the electrophysiological study. (WDR - Wide-dynamic range; NS nociceptive-specific).

Spontaneous activity of the RVM neurons prior to stimulation is represented in [Figure 15](#page-49-0). Statistical analysis showed an effect of ovariectomy on the spontaneous activity of NS Off-like cells ([Table 11](#page-49-1)), and further multiple comparisons showed an increase in CCI/SHAM animals when compared to CCI/OVX animals (P=0.044). No differences were observed between the remaining RVM cell types.

Figure 15. Spontaneous response of RVM neurons before noxious mechanical stimulation of the tail (pinching with a surgical clamp). Differences were observed on NS Off-like cells, with CCI/SHAM animals displaying an increased spotaneous activity than CCI/OVX females. Results are presented as mean ± SEM. *P<0.05 (t-test with a Bonferroni correction for multiple comparisons). (WDR - Wide-dynamic range; NS - nociceptive-specific).

Table 11. Results of the statistical analysis of the differences in the spontaneous activity of RVM neurons before noxious mechanical stimulation of the tail (pinching with a surgical clamp) between female experimental groups. *P<0.05. (WDR - Wide-dynamic range; NS - nociceptive-specific; CCI - chronic constriction injury).

After noxious mechanical stimulation ([Figure 16](#page-50-0)), statistical analysis showed CCI increased the response of WDR On-like cells, as well as an interaction between CCI and ovariectomy on the evoked response of NS Off-like cells ([Table 12](#page-50-1)). However, multiple comparisons showed no differences between female experimental groups for all cell types. Neuropathic males showed a decreased response of WDR $(CCI/SHAM: t_{46}=3.57, P<0.001; CCI/OVX: t_{47}=2.66, P=0.011)$ and NS On-like cells $(CCI/SHAM: t_{40}=3.72,$ $P<0.001$; CCI/OVX: t₃₉=3.58, P<0.001), in comparison with both CCI/SHAM and CCI/OVX females. CCI_M also showed a decreased response of WDR Off-like cells comparing with CCI/SHAM ($t_{41}=2.78$, P=0.012).

Figure 16. Evoked response of RVM neurons after noxious mechanical stimulation of the tail (pinching with a surgical clamp). No significant differences were observed between experimental groups. (WDR - Wide-dynamic range; NS - nociceptive-specific). Results are presented as mean \pm SEM. # P<0.05 and ### P<0.001 (unpaired ttest with Welch's correction).

Table 12. Results of the statistical analysis of the differences in the evoked response of RVM neurons to noxious mechanical stimulation of the tail (pinching with a surgical clamp) between female experimental groups. *P<0.05. (WDR - Wide-dynamic range; NS - nociceptive-specific; CCI - chronic constriction injury).

4.5.3. Cold stimulation

Regarding innocuous thermal stimulation, RVM neuronal activity was recorded when acetone was applied to the plantar skin of the left hindpaw. The classification and the total number of neurons recorded is described in [Table 13](#page-51-0).

Table 13. Total number of RVM On-, Off- and Neutral-like cells to innocuous thermal stimulation (acetone applied to the plantar skin of the left hindpaw), recorded in the electrophysiological study. (WDR - Wide-dynamic range; NS - nociceptive-specific).

Prior to stimulation with acetone, the spontaneous activity of RVM neurons is represented in [Figure 17](#page-51-1).

Figure 17. Spontaneous activity of RVM neurons before cold stimulation (acetone applied to the plantar skin of the left hindpaw). CCI /SHAM animals showed an increased spontaneous activity than SHAM/SHAM animals. Results are presented as mean \pm SEM. *P<0.05 (t-test with a Bonferroni correction for multiple comparisons). (WDR - Wide-dynamic range; NS - nociceptive-specific).

No differences were observed in the spontaneous activity of WDR neurons. Regarding NS neurons, the spontaneous activity of NS On-like cells was increased after de induction of CCI ([Table 14](#page-52-0)), and multiple comparisons showed increased spontaneous activity in CCI/SHAM compared with SHAM/OVX animals (P=0.018). Also, an interaction between CCI and ovariectomy was observed in NS Off-like cells, and post hoc multiple comparisons showed increased spontaneous activity in SHAM/OVX, in comparison with SHAM/SHAM animals (P=0.048). Interestingly, no differences were found when comparing neuropathic males and females.

Table 14. Results of the statistical analysis of the differences in the spontaneous activity of RVM neurons before cold stimulation between female experimental groups. ** P<0.01. (WDR - Wide-dynamic range; NS - nociceptivespecific; CCI - chronic constriction injury).

	Interaction	Ovariectomy	CCI
WDR On-like cells	$F_{1,29}$ =0.44, P=0.51, η _s ² =0.011	$F_{1,29} = 1.01$, P=0.32, $\eta_{p}^2 = 0.024$	$F_{1,29} = 1.62$, P=0.21, $\eta_{p}^2 = 0.038$
WDR Off-like cells	$F_{1,31} = 1.98$, P=0.17, η_{ρ} ² =0.058	$F_{1,31}$ =0.65, P=0.42, η_{s} ² =0.019	$F_{1,31}$ =0.20, P=0.66, η_{ρ} ² =0.006
NS On-like cells	F_{127} =0.11, P=0.008, n _s ² =0.003	$F_{127} = 3.88$, P=0.059, n ² =0.099	$F_{1,27} = 8.24$, P=0.008 ^{**} , $\eta_{p}^2 = 0.21$
NS Off-like cells	$F_{1.25} = 8.34$, P=0.008 ^{**} , $\eta_e^2 = 0.21$	$F_{125} = 3.10$, P=0.091, n $^2 = 0.076$	$F_{1.25} = 4.06$, P=0.055, $\eta_{\text{B}}^2 = 0.10$

After stimulation with acetone ([Figure 18](#page-52-1)), CCI independently decreased the response of WDR On-like neurons (Table 15), and multiple comparisons showed differences between SHAM/OVX animals and both CCI/SHAM (P=0.018) and CCI/OVX neuropathic females (P=0.028). No differences were observed between experimental groups regarding NS and WDR Off-like neurons. Identically, comparisons between male and female CCI groups showed no statistically significant differences.

Figure 18. Evoked response of RVM neurons after cold stimulation (acetone applied to the plantar skin of the left hindpaw). CCI females (CCI/SHAM and CCI/OVX) showed a decreased response of WDR On-like cells, in comparison with SHAM/OVX. Results are presented as mean \pm SEM. *P<0.05 (t-test with a Bonferroni correction for multiple comparisons. (WDR - Wide-dynamic range; NS - nociceptive-specific).

Table 15. Results of the statistical analysis of the differences in the evoked activity of RVM neurons after cold stimulation (acetone applied to the plantar skin of the hindpaw) between female experimental groups. (WDR - Widedynamic range; NS - nociceptive-specific; CCI - chronic constriction injury).

4.6. Histopathological analysis of the sciatic nerve

At the end of the experimental period, a section of the ipsilateral sciatic nerve was used for histopathological assessment (Figure 19).

Control animals (SHAM/SHAM) showed a clear organization of nerve fibres and absence of infiltrating cells. Ovariectomy alone caused no histopathological alterations to the sciatic nerves, with no differences between SHAM/SHAM and SHAM/OVX females. On the other hand, differences were observed between CCI and SHAM females. Longitudinal sections of the sciatic nerve of CCI animals showed a loss of organization of nerve fibres, with the presence of degraded myelin sheath as well as several infiltrating inflammatory cells. This loss of fibre organization was also possible to observe in transversal sections, as well as, the presence of inflammatory cells. Interestingly, these infiltrates were more abundant on the sciatic nerves of CCI/OVX females.

Figure 19. Histopathological analysis of the left sciatic nerve. Representative micrographs of longitudinal (Left) and transverse (Right) cuts for each female experimental group. It is possible to observe the presence of degraded myelin sheath (*) as well as infiltration of inflammatory cells (arrows) in the sciatic nerve of CCI females. Images were obtained using an Olympus BX61 brightfield microscope, coupled to an Olympus DP70 camera at the original objective magnification of 20X

In this work, the induction of CCI in female rats impaired body weight gain and lead to the development of mechanical and cold allodynia from week 1 post-CCI surgery onwards. Importantly, and contrary to what is described for males, CCI in female Wistar Han rats did not induce the development of emotional impairments, namely anxiety- or depressive-like behaviours. Interestingly, ovariectomy induced anhedonic-like behaviour with CCI/OVX females also displaying locomotor impairments. The electrophysiological results partly support our behavioural data to the extent that increased mechanical sensitivity in females and nociceptive recovery of CCI males was concomitant to changes in the neuronal activity of the RVM WDR and NS neurons. Nociceptive changes in CCI females were also accompanied by histopathological alterations of the sciatic nerve architecture.

5.1. Technical considerations

5.1.1. Animal model

The study of chronic pain depends on the use of animal models to evaluate their sensory and psychological intricacies. As mentioned previously, the CCI model of neuropathic pain⁹¹ mimics the symptoms of chronic nerve compression or entrapment in various clinical conditions, comprising both the inflammatory and neuropathic components. This model distinguishes itself from other neuropathic pain models through the display of signs of spontaneous pain such as limping, limb guarding, excessive licking and avoidance of weight bearing on the injured limb. Notwithstanding its various advantages, the CCI model also has its demerits, such as variation in the tension of the ligatures throughout the experimental period and during induction, leading to inter-individual variability, even with the same experimenter. Another disadvantage of this model is the development of autotomy in some animals, which always progressed from the claws to the root of the toes. The root of the problem is probably the excessive trimming of the claws, as dysesthesia would damage the hindpaw randomly.

By using OVX animals, we were able to control the potential effect of hormonal depletion upon nociceptive and emotional behaviours, as well as upon neuronal activity. For the ovariectomy procedure, we opted to perform a dorsal and bilateral technique²²⁷. Primarily, an ovariectomy is preferred over an ovariohysterectomy as the manipulation of the uterus can lead to more surgical complications²³⁵. In

42

rodents, the dorsal approach is simple and avoids the need to manipulate and/or injury the gastrointestinal tract. In addition, as only a single incision to the skin is needed to access both ovaries, this approached is considered less invasive. It is, however, important to consider that performing a gonadectomy disrupts the normal feedback loop between gonadal hormones and the anterior pituitary and hypothalamus^{236,237}. This leaves the animals in a prolonged state of altered circulating hormones causing a variety of uncontrolled consequences. The use of randomly cycling female rats did not allow to evaluate the role of gonadal hormone fluctuations on nociceptive and emotional processing. Nonetheless, we did not observe differences in variability of our results between gonadally intact and ovariectomized females.

The present work focused on the use of female experimental animals, as this model has been thoroughly characterized in male rats. However, a group of male CCI animals (CCI_M) was also included to allow comparisons between neuropathic females and males, as well as for validation of the experimental outcomes as literature on CCI males is extensive.

Regarding animal welfare, we evaluated body weight change throughout the experimental procedures. In females, ovariectomy was shown to increase the body weight of the animals, consistent with the previous reports²³⁸. The loss of ovarian function in rats has been known to increase daily food intake, body mass and body fat mass, which can be reversed by treatment with estradiol²³⁹. Interestingly, we observed this model of neuropathic pain arrested weight gain on female rats, which is not observed in males. However, our results are validated since as in other reports, CCI males continued to increase body weight despite the induction of CCI177,196.

5.1.2. Behavioural assessment of nociception

One of the greatest difficulties in preclinical pain research is to find behavioural outcomes that properly evaluate the pain experienced by rodents. Since animals cannot self-report, the assessment of the extent and severity of pain is another major challenge in all pre-clinical models. Therefore, a battery of behavioural tests has been developed for the evaluation of nociception, with most of these tests, such as the herein used, measuring the response to a thermal or mechanical stimulus²⁴⁰. In this work, we assessed mechanical and cold allodynia using the von Frey and acetone tests, respectively. There are other behavioural measures of pain, focused on its consequences rather than behavioural responses, that may also reflect patients' symptomatology. These include the assessment of (i) adaptive postural changes, which contribute to physical disability; (ii) spontaneous behaviours, evaluating the impact of pain on daily

activities and (ii) motivational aspects, as motivation to pain relief and avoidance of painful situations²⁴¹. These evaluations are however very time-consuming and difficult to evaluate quantitatively, while quantitative measures of allodynia/hyperalgesia are easily quantified, increase repeatability and reproducibility of the results and are present in clinical pain.

Mechanical allodynia is a high constraint symptom in chronic pain. In animal models, mechanical allodynia is often addressed using von Frey filaments applied to the plantar surface of the hindpaws²²⁸. The more common methodologies to determine mechanical sensitivity are the "ascending stimulus", "percent response" and "up-down" methods. While the "ascending stimulus" method avoids excessive stimulation of the hindpaw, it provides only an estimate of the withdrawal threshold. On the other way around, in the percent response" method the number of stimuli per test is significantly higher than other methodologies. In this work, the "up-down" method was used to calculate the mechanical force required to elicit a paw withdrawal response in 50% of animals (50% threshold)228,242. A disadvantage of this test is that the number of stimuli per animal is variable and requires repeated time-intensive measurements. This repeated stimulation can cause the animals to develop sensitization or learnt responses. Nevertheless, this test has the advantage of unrestrained assessment, being, therefore, less stressinducing.

In rodents, a variety of tests have also been developed for the evaluation of cold allodynia such as immersion in cold water^{243,244}, water-alcohol bath²⁴³, ethyl chloride spray²⁴⁵, cold plate²⁴⁶ and the herein used acetone test²⁴⁷. While these tests are easily performed and could provide more accurate quantification of cold sensitivity, they require specific equipment. In the acetone test, a small drop of acetone is applied to the plantar skin of the hindpaw. The great advantage of this test is the use of the same apparatus as for the von Frey test and acetone's low cost and availability. The acetone test incorporates a complex, multimodal stimulus comprising simultaneously active cold, mechanical, and chemical components. To avoid a mechanical stimulus, the acetone was applied without touching the hindpaw with the pipette. Probably, this leads to the activation of multiple types of nociceptors of the affected hindpaw²⁴⁷. Another potential disadvantage of this test is the reaction of the animals to the smell of acetone, which can lead to agitation and false responses. To avoid it, we habituated the animals to the smell of acetone prior to performing the test.

5.1.3. Behavioural paradigms of emotional behaviour

Since emotional disorders can only be evaluated indirectly, we then refer to them as anxiety-like and depressive-like behaviours. These can be assessed using a wide range of behavioural testing paradigms, which assume that rodents and humans share, to some extent, innate behaviours when faced with new challenges. However, it is necessary to take into consideration that each individual test assesses only a fraction of the emotional profile of the animal.

For the evaluation of anxiety-like behaviours, the OF and the EPM tests were performed, two of the most commonly used tests in rats²⁴⁸. The OF is an exploration test, as it is based on the animals' inner conflict between approaching or avoiding novel environments²⁴⁹. One of the greatest advantages of this tests is that it allows the analysis of both locomotion and anxiety: locomotion is evaluated through the total distance travelled by the animals, and the time spent in the centre of the arena is a measure of anxietylike behaviour. Also, through an infrared beam system, it is largely automated and with no interexperimenter variability on its analysis. Despite its advantages, it has been shown that OF activity can be influenced by several factors such as the species, strain and sex of the animals as well as the equipment used²⁴⁹. Similar to the OF test, the EPM is a very simple method to assess anxiety-like behaviour in rodents. This test is easy to quantify and does not comprise the presentation of noxious stimuli, relying only on the rodent's avoidance of open spaces and heights. It is important to take into consideration that the results on both these tests are influenced by habituation, resulting in the reduction of activity, so this test was only performed once. Also, while the OF test is more exploratory-based and suitable for locomotor measurements, the EPM is more suitable for anxiety testing. As some behavioural tests depend on the locomotor ability of the animals, the basal locomotor ability (without effort) of our animals was assessed in the OF test by evaluating the velocity and total distance travelled.

To evaluate depressive-like behaviours, we chose to assess two components of depression: anhedonia (SPT) and learned helplessness (FST). Regarding the SPT²⁵⁰, this is an "effortless" test as it does not imply intense physical activity in order to gain access to the sweet solution. Also, our protocol does not require food and/or water deprivation prior to the test, reduces its duration and consequently the anxiety induced by isolation from the respective cage mates. Nonetheless, as this test lasts only a few hours, factors such as circadian rhythm and interindividual differences in the pattern and amount of liquid intake can influence the results. The analysis of anhedonia in rodents should be approached with caution, a reduced sucrose preference is considered to share some analogy with anhedonia and depressive behaviour in humans²⁵¹ and thus, SPT is widely used in chronic stress studies. Regarding the FST¹⁹², a great advantage

of this test is that, through filling the tank in such depth that the animals cannot reach the ground, it allows the distinction between active behaviours (swimming/climbing), which is known to be differentially due to different serotonergic and noradrenergic mechanisms188,189 . However, repeated testing and consequent learning processes can influence the results, as the animals recognize there is no escape from that situation and, for that reason, we opted to perform the test only once. Also, as this test is sensible to variation between observers, video recordings were analysed by two independent researchers.

It is important to point out all our emotional behavioural tests were performed on week 5 post-CCI surgery, while most studies perform anxiodepressive testing on week 4. Our choice was first based on the fact that emotional impairments, namely depressive-like behaviour is a slow-setting condition, taking several weeks to fully develop. Secondly, the development of nociceptive impairments was delayed and less extensive in females when compared to male animals. Thirdly, while the chromic cat gut used to constrict the sciatic nerve is known to deteriorate from day 28 onwards, several reports showed the presence of emotional impairments in male animals at later dates.

5.1.4. Electrophysiological recordings

Some consideration must be given to the advantages and disadvantages of the type of electrophysiological recordings used, namely a single cell recording technique. This technique allows the recording of a few cells simultaneously in every session, allowing the differentiation of functionally heterogeneous populations of neurons in the same area. This is of interest when recording the RVM, as it is comprised of functionally different cell types. As well, this allows the comparison between basal and neuronal activity evoked by several innocuous and noxious stimuli. After categorizing the recorded cells between NS and WDR, based on their response to noxious and innocuous stimuli, these cells were subdivided based on changes in their discharge rate (10% more or less in relation to spontaneous discharge rate²³²) after peripheral stimulation. A disadvantage of this classification is that, in cells with low spontaneous firing rates, a very small change is enough to classify the cell as responsive. For this reason, we considered a minimum variation of 0.45 spikes.s¹ to classify a cell as responsive.

Despite the advantages above mentioned, the results obtained must be analysed carefully. It is important to note that the experimental procedures were performed in animals under anaesthesia, which may interfere with brain function and bias the results due to possible effects of the sedation. Also, in large and heterogeneous areas, the small population of neurons analysed may not be a full representative of the entire area of interest. To partially overcome this constraint, we recorded as many neurons as possible

46

per session and at different depths. Concerning the experimental procedure, maintaining an anaesthetized state using sodium pentobarbital depended on a regular intraperitoneal injection, compromising the stability of the anaesthesia level and, consequently, the firing activity of the recorded cells. Thus, in addition to verifying the dilation of the pupils, the general muscle tone and responses to noxious pinching every 15 min, we also used spontaneous activity to control for variation in anaesthesia level. Finally, pentobarbitone has a narrow safety margin for anaesthesia, and mortality can be high – leading to a smaller sample size and fewer neurons being recorded in some animals.

5.2. Development of neuropathic pain

The hallmarks of neuropathic pain comprise the development of spontaneous pain - pain arising without stimulus -, allodynia - abnormal responses to non-painful stimuli – and hyperalgesia – exacerbated response to painful stimuli²⁵³.

Spontaneous pain is particularly difficult to measure in rodents but since it was not within the scope of this work, no attempt at performing direct measures was made. Nevertheless, CCI animals displayed signs of limping and guarding the affected hindpaw, behaviours which are in line with those described by Bennet and Xie⁹¹, thus supporting the CCI model was effectively induced in our animals. Also, as described earlier⁹¹, autotomy was sometimes observed, with gnawed claw tips, leading to the exclusion of some CCI animals from the further nociceptive evaluation. This behaviour is present on conditions that cause anaesthesia dolorosa and was proposed to be triggered by abnormal afferent signaling generated in the ligation site²⁵⁴. Importantly, no differences were observed between neuropathic males and females regarding this parameter.

Neuropathy caused by CCI surgery led to the development of mechanical and cold allodynia, in both CCI/SHAM and CCI/OVX females, starting from week 1. While ovariectomy solely did not lead to detectable nociceptive abnormalities, gonadally intact and ovariectomized females with CCI showed differences in the development of mechanical allodynia. Whereas CCI/OVX females developed mechanical allodynia prior to CCI/SHAM, gonadally intact females showed increasingly lower withdrawal thresholds throughout the weeks while ovariectomy stagnated the development of mechanical allodynia. Some studies have shown alterations in sensitivity to nociceptive stimuli following OVX. A recent study by Li and colleagues²³⁸ showed the development of mechanical allodynia 5 weeks after OVX, using the von Frey test, however, in this same study, no differences were seen between intact and ovariectomized females one-week post-CCI, which we observed in this work. As this assessment was performed only once after CCI induction, it may not reflect the time-dependent development of mechanical allodynia observed herein. Altogether, these results point to a role of gonadal hormones not only on the early development of CCI-induced allodynia but also impacting the further maintenance of nociceptive symptoms.

As expected, CCI-induced neuropathy led to the development of mechanical and cold allodynia, in both CCI/OVX and CCI/SHAM females. While ovariectomy by itself did not lead to nociceptive abnormalities, CCI/SHAM and CCI/OVX females displayed differences in the development of mechanical allodynia, with CCI/OVX females showing symptoms two weeks prior to CCI/SHAM. In addition, while mechanical allodynia is gradually worsened in CCI/SHAM females, CCI/OVX partially recovered in the last experimental week. Some studies have shown alterations in sensitivity to nociceptive stimuli following O^V . A recent study by Li and colleagues²³⁸ showed the development of mechanical allodynia 5 weeks after OVX, using the von Frey test, however, in this same study and contrary to our results, no differences were seen between intact and ovariectomized females one-week post-CCI. As Li's assessment was performed only once after CCI induction, it may not reflect the time-dependent development of mechanical allodynia observed herein. These results point to a role of gonadal hormones not only on the early development of CCI-induced allodynia but also in the maintenance of nociceptive symptoms.

The establishment of CCI in our females is further validated by results from the CCI_M group. The development of mechanical and cold allodynia in CCI males followed a similar pattern to that of females. However, despite displaying lower withdrawal thresholds earlier when compared to CCI females at weeks 1 and 2, recovery signs started at week 3 and full recovery was observed at week 5. These results are in clear contrast to those of CCI females as these did not recover to baseline values, displaying nociceptive responses until week 5. This sex-specific difference is in accordance with what was observed by Tall and colleagues²²¹, where neither ovariectomized nor intact female rats recovered after the induction of neuropathic pain, contrary to what's observed in males by day 35 post-CCI surgery. Another work from Vacca *et al.* 224 also observed in mice that, while males showed a gradual decrease of allodynic responses, females showed no recovery throughout 121 days of CCI. This data supports slower regenerative processes resulting from the injury in female animals when compared with males.

Altogether, our results highlight the existence of sex-differences in the CCI model, not only on the development and maintenance of neuropathic pain but also on general welfare measures such as body weight, as discussed in the technical considerations section. Importantly, we show an impact of ovarian hormones on body weight as well as on the development of mechanical allodynia on CCI females which should be further investigated to verify if it interferes with the efficiency of pain management therapies.

While CCI by itself did not affect the locomotor ability of females, when associated with OVX it highlighted the development of locomotor impairments in CCI/OVX females translated in a significant decrease in the total distance travelled in the OF, probably due to decreased velocity. Accordingly, while the locomotor performance of CCI/SHAM females was identical to that of males, the total distance travelled of CCI/OVX females was significantly lower than that of CCI males. Except for CCI/OVX females, our data is in accordance with literature showing no motor dysfunctions in neuropathic animals¹⁶⁹. However, a work by Li and colleagues²³⁸ showed ovariectomy reduced the distance travelled in the OF thus support our CCI/OVX data. These results call to the attention that concomitant CCI and ovariectomy impair the locomotor abilities of female animals and these must be considered when performing locomotiondependent behavioural tests. Additionally, it is important to note that the performance of CCI males and CCI/OVX females cannot be considered comparable.

5.3. Neuropathic pain and the development of mood disorders

In male animals, the development of emotional impairments in CCI has been thoroughly described, both regarding anxiety- and depressive-like behaviour⁶¹, as shown in **Section [1.7](#page-21-0)**. The opposite is true for female rats thus being of great interest to uncover sex differences on the comorbidity between experimental neuropathic pain and emotional impairments.

In terms of anxiety-like behaviour, our results were not consistent among tests since the increased defecation of CCI females in the OF suggested the development of anxiety-like behaviour, but this observation was not supported by the remaining OF and EPM data. Although negative, our results were not completely unexpected as only one study¹⁷⁵ (**Section 1.7.1**) reported the development of anxiety-like behaviour, using the OF and EPM tests, on right operated intact and ovariectomized female mice on day 26. On the other hand, CCl_M performance indicates the development of anxiety-like behaviour in these animals. The number of *faecal boli* left in the OF arena by CCI_M was similar to that of CCI females, and these animals spent considerably less time in the EPM open arms than their female counterparts. The measurement of *faecal boli* is not validated across laboratories and is usually only used to support other anxiogenic measures. So, while it is interesting that results between CCI females and males were identical, it is not enough to establish an anxious-like phenotype.

One possible reason for CCI females not displaying an anxious-like phenotype is the time point chosen to evaluate. In fact, it has been demonstrated in the cuff model of chronic neuropathic pain²⁵⁵ that anxietylike behaviour is present in earlier stages, while depressive-like behaviour develops later on. Accordingly,

a study using the sciatic nerve ligation (SNL) of experimental neuropathic pain²⁵⁶ showed a decrease in the percentage of time spent in the open arms of the EPM on day 20 post neuropathic pain induction.

Concerning ovariectomy, a recent study by Puga-Olguín et al.²⁵⁷ showed the development of anxiety-like behaviour, using the EPM, three weeks after ovariectomy through a decrease in the time spent in open arms. Thus, since we tested our animals in week 5, it is possible we lost this window of opportunity. On the other hand, the intensity and timeline of mechanical allodynia settlement in females, especially in CCI/SHAM females, was slower when compared with CCI_{M} , and, if so, it is possible that more time was needed for the development of anxiety-like behaviour in our CCI females. This assumption is also partly supported by our OVX females data as literature shows only long-term ovariectomy leads to an anxiogenic state in the OF test in female rats and mice²⁵⁸⁻²⁶⁰.

In what regards depressive-like behaviours, two different components were assessed: anhedonia through the SPT and learned helplessness in the FST. Identically to what was observed for anxiety, the induction of CCI did not lead to the development of depressive-like behaviour in females.

Considering CCI females still displayed mechanical and cold allodynia at the end of the experimental period, SPT results in CCI females were rather unexpected. In a study using chemically induced experimental gastritis²⁶¹, female, but not male, Sprague-Dawley rats displayed decreased sucrose preference. However, there is a strong inflammatory component associated with gastritis which might account for the behavioural differences in females. Our data are nonetheless in accordance with a study using the SNL²⁶² where females also did not decrease sucrose preference. Comparability between works is however limited since not only the neuropathic pain model used are different, but one evaluation was performed on day 15 post SNL induction and ours much later. As argued previously in relation to anxiety, it is also possible we missed the time window to detect it, thus both an earlier (3-4 weeks) and a later (8- 12 weeks) timepoints should be investigated in future studies. Nonetheless, the SPT results in our OVX females are in accordance with previous reports showing the development of anhedonic-like behaviour on day 28²⁶³, 5 weeks²⁶⁴ and 4 months post-OVX induction²⁶⁵.

Interestingly, CCl_M displayed anhedonic-like behaviour but not learned helplessness. Of the four works using SPT (Section 1.7.2), one in mice showed anhedonic-like behaviour on right side operated animals between weeks 4 and 10 while in rats¹⁷⁰, one study using right-operated Sprague-Dawley rats showed no anhedonia¹⁶⁸ and two using right side operated Wistar Han rats showed anhedonic-like behaviour on days 9, 16 and 28^{208,215}. Interestingly, all these studies were performed on right side operated animals, while in this work CCI was induced on the left. Again, data from CCl_M adds to the literature, showing that while mechanical and cold allodynia was apparently resolved in males, these animals still display anhedoniclike behaviour.

Lack of significant differences between experimental groups in the FST was surprising, especially for the CCI_M group since these animals displayed anhedonic-like behaviour. Concerning CCI females, our results contrast with data from the study using SNL²⁶² where neuropathic females displayed increased immobility and decreased time spent swimming in the FST. In addition, another work²⁵⁶ showed similar results but on day 20. Yet, these differences might again not only be related to the use of different experimental models of neuropathic pain but also different testing time points, 15 and 20 days vs. 5 weeks.

Contrary to what was observed for anhedonic-like behaviour, ovariectomy did not induce the development of behavioural despair. The literature on this subject is contradictory, with different results being reported depending on the strain and the testing timepoints chosen. In female Wistar rats, ovariectomy was shown to increase immobility at week 1, but not at weeks 3 and 12^{238} or after 15 months²⁶⁶ post-surgery, which is in accordance with our results. However, increased immobility was observed on recent studies, namely 6 weeks after ovariectomy²⁵⁷ in Wistar Han rats and in Sprague-Dawley rats at week 5²³⁸. Thus, and as proposed before, for a better understanding of the effect of ovariectomy on depressive-like behaviours, further studies should include different post-ovariectomy periods to assess the possibility of a timedependent effect of this behaviour.

As male animals were shown to develop learned helplessness following CCI (Section 1.7.2), our CCI_M results come as unexpected. Data from previous works on males show learned helplessness was observed up to week 6 on Sprague-Dawley rats¹⁹⁸ and week 5 on Wistar Han rats¹⁶⁷. It is possible the amelioration of mechanical and cold allodynia from week 3 onwards prevented the development of learned helplessness in these animals. In fact, a study²⁰⁸ using right side operated Wistar Han rats showed increased immobility and decreased time spent climbing in the FST on day 28 post CCI-induction but paw-withdrawal threshold remained low. Identically, in another two studies left side operated Wistar Han rats also display increased immobility time in the FST167,196 and decreased time spent climbing¹⁹⁶ on day 28 but again paw-withdrawal threshold in these animals remained low.

Altogether, our results highlight the existence of sex differences in the development of anxiety- and depressive-like behaviours after CCI-induced neuropathy, as well as an impact of hormonal withdrawal on anhedonic behaviour of ovariectomized female rats.

5.4. CCI-induced changes in RVM neuronal activity

In this work, we assessed whether RVM neuronal activity was affected by CCI-induced neuropathic pain or ovariectomy. The RVM is an area involved in descending modulation of pain, through a balance between the activity of RVM pronociceptive On- and antinociceptive Off-cells. As clinical studies show hypersensitivity to cold and mechanical stimulation are hallmarks of neuropathic injuries²⁶⁷, when assessing the response of RVM cells to peripheral stimulation we focused on these stimuli.

While the spontaneous activity of RVM WDR and NS cells was not increased prior to VF stimulation in females, we observed in CCI males an increase in the spontaneous activity of NS Off-like cells. This increase in the tonic activity of NS RVM OFF-like neurons can, at least partly, be correlated to the recovery observed in behavioural mechanical allodynia due to the antinociceptive role of these cells. Interestingly, during VF stimulation, the response of WDR Off-like cells in the CCI/OVX group was significantly increased which, considering their antinociceptive role, supports the increased in behavioural mechanical allodynia threshold in comparison with CCI/SHAM females on week 5. CCI-induction per se did not alter RVM cells response to von Frey stimulation, which is in accordance with a study by Heinricher *et al.*²⁶⁸ where, in the SNL model, no differences were observed between Sham and ligated animals during von Frey stimulation.

Regarding tail pinch stimulation, results are more difficult to interpret because the activity of both RVM On- and Off-like cells is enhanced. Nonetheless, our results suggest the heightened response of pronociceptive WDR RVM On-like cells to noxious stimulation is matched by an attempt to counteract it by WDR RVM Off-like cells in CCI/OVX females. Again, this falls in line with the increase in behavioural mechanical allodynia threshold in comparison with CCI/SHAM females on week 5. The increased response of pronociceptive female On-like cells when compared to that of neuropathic males, suggests increased descending pronociception and could partly explain the differences in behavioural mechanical allodynia experienced between female and male CCI animals at the end of the experimental period.

While our results suggest a profound alteration of RVM noxious stimuli processing in CCI females, with increased descending pronociception when compared with CCI males, it is important to remember this noxious stimulation was applied to the tail and not the affected hindpaw.

Despite the fact that CCI did not alter the spontaneous activity of pinch-responsive neurons, previous studies using other neuropathic²⁶⁹ and inflammatory²⁷⁰ chronic pain models showed altered spontaneous activity of RVM On/Off-cells. In a diabetic neuropathy model²⁶⁹, an increase in the spontaneous activity of On-cells, as well as a decrease in Off-cells, was demonstrated. However, in an arthritis model²⁷⁰, both RVM

On- and Off-cells increased spontaneous activity, similarly to our results. These findings indicate the recruitment of RVM On- and Off-cells differs significantly between neuropathic and inflammatory chronic pain models.

Regarding cold stimulation, the increase in the spontaneous activity of NS RVM On-like neurons in CCI animals suggests enhanced activation of pronociceptive descending pathways with a potential counteracting effect of NS RVM Off-cells in CCI/OVX females. Surprisingly, especially bearing in mind the behavioural cold allodynia data, during stimulation with acetone the response of WDR RVM On-like cells is decreased in CCI females. Our results also in opposition to those reported by a previous study using the SNI model of neuropathic pain²⁶⁹ and showing an increase in cold-evoked responses of RVM On-like cells at week 8.

Overall the activity of RVM pain modulatory neurones partially supports our behavioural observations, except in what concerns cold allodynia. Differences in mechanical allodynia between CCI/SHAM and CCI/OVX animals are concomitant with subtle changes in both the spontaneous and evoked activity of mainly RVM WDR cells. Differences between the activity of RVM neurones of CCI females and males suggest enhanced activity of descending pronociceptive pathways in the former. Nociceptive recovery of CCI males appears to be associated with an overall concomitant increased in the spontaneous activity of RVM NS antinociceptive neurones and a decrease in the evoked activity of RVM WDR and NS pronociceptive cells.

Additionally, studies point to a possible role of Neutral cells in nociceptive modulation 271 despite their lack of response to acute peripheral noxious stimulation. The latter were not assessed in this work however they emerge as a potential target of future studies.

In sum, these results suggest an increase in descending pronociception in female rats by both CCI and ovariectomy, modulated by the RVM. This pronociceptive shift is known to facilitate nociceptive signaling and could explain the exacerbated mechanical allodynia seen in females, in comparison with males.

5.5. Histological evaluation of the sciatic nerve

We evaluated whether CCI induced histopathological changes in the structure of the sciatic nerve. During sampling of the nerve, we observed the ligated region was opaque in a dull yellow color, and through H&E staining, we observed that nerve constriction led to degraded myelin sheets and inflammation. These are common consequences of peripheral nerve injury, confirming the proper induction of the CCI model, as well as the inflammatory response induced by the chromic gut suture. This same CCI-induced pathology

was also observed in previous studies, from nerve derangement to inflammation on the affected site^{272,273}. Interestingly, we observed an increase in inflammatory infiltrates in CCI/OVX females which could be explained by the more pro-inflammatory state induced by ovariectomy²⁷⁴.

Nonetheless, our analysis had its setbacks as some of the sciatic nerves were damaged throughout processing and prevented a clear visualization of the nerve structure. As well, the absence of data from males due to time-constraints hinders comparisons between neuropathic males and females. It would be of interest in the future for these samples to be assessed by a senior neuropathologist to confirm our results. Future studies could also include the evaluation of demyelination in the sciatic nerve of neuropathic animals through more thorough histological and techniques.

Chapter 6. Concluding remarks and future perspectives

In the present thesis, using a behavioural and electrophysiological approach, we demonstrated: (i) CCI lead to the development of both mechanical and cold allodynia in female rats, that, contrary to what is observed in males, do not show signs of recovery at 5 weeks post-CCI surgery; (ii) CCI did not induce the development of anxiety or depressive-like behaviours in females; (iii) ovariectomy induced anhedonic-like behaviour, and locomotor impairments in CCI females; (iv) neuropathic males perform differently from females on anxiety- and depressive-like behavioural paradigms and (v) CCI and ovariectomy induced alterations in the neuronal activity of the RVM, with increased descending pronociception on week 6 post-CCI surgery.

Our results highlight the existence of sex differences in the development and maintenance of CCI-induced neuropathic pain as well as emotional comorbidities. Thus, our data challenge the assumption the use of male subjects on basic research is equivalent to that of females and straightforwardly translatable to women. Future studies should try to elucidate the mechanisms underlying these sex differences, and how this impacts the efficiency of chronic pain management therapies.

The observed differences between gonadally intact and ovariectomized females further confirm the role of gonadal hormones in this neuropathic pain-emotions comorbidity. Further work should investigate hormonal replacement therapy to determine which and how gonadal hormones are involved in these alterations. In addition, it would be of interest to assess if ovariectomy would differentially affect the development of emotional impairments if performed after the induction of peripheral neuropathy.

It remains to be studied how CCI-induced changes in the response properties of RVM neurons observed could be altering supraspinal processing or the response of spinal dorsal horn neurons after peripheral neuropathy, as well as if these can be modulated through pharmacological intervention. Also, possible cellular and structural changes could be associated with the development of chronic neuropathic pain and further studies could include the stereological analysis of the RVM as well as other areas involved in pain and emotional modulation.

55

Chapter 7. References

- 1. Neugebauer, V., Galhardo, V., Maione, S. & Mackey, S. C. Forebrain pain mechanisms. Brain Res. Rev. 60, 226–242 (2009).
- 2. Baliki, M. N. & Apkarian, A. V. Nociception, Pain, Negative Moods, and Behavior Selection. Neuron 87, 474–491 (2015).
- 3. Brown, A. G. & Brown, A. G. in *Organ. Spinal Cord* 1–12 (Springer London, 2011).
- 4. Basbaum, A. I., Bautista, D. M., Scherrer, G. & Julius, D. Cellular and Molecular Mechanisms of Pain. *Cell* 139, 267-284 (2009).
- 5. Djouhri, L. & Lawson, S. N. Aβ-fiber nociceptive primary afferent neurons: A review of incidence and properties in relation to other afferent A-fiber neurons in mammals. Brain Res. Rev. 46, 131-145 (2004).
- 6. D'Mello, R. & Dickenson, A. H. Spinal cord mechanisms of pain. Br. J. Anaesth. 101, 8-16 (2008).
- 7. Dubin, A. E. & Patapoutian, A. Nociceptors: the sensors of the pain pathway. J. Clin. Invest. 120, 3760–3772 (2010).
- 8. Julius, D. & Basbaum, A. I. Molecular mechanisms of nociception. *Nature* **413,** 203–210 (2001).
- 9. Todd, A. J. Neuronal circuitry for pain processing in the dorsal horn. Nat. Rev. Neurosci. 11, 823– 36 (2010).
- 10. Kandel, E. R., Schwartz, J. H., Jessel, T. M., Siegelbaum, S. A. & Hudspeth, A. J. Principles of neural science. (McGraw-Hill QDR, 2013).
- 11. Renn, C. L. C. L. & Dorsey, S. G. S. G. The physiology and processing of pain: a review. AACN Clin. Issues 16, 277-90; quiz 413-5 (2005).
- 12. Bourne, S., Machado, A. G. & Nagel, S. J. Basic anatomy and physiology of pain pathways. Neurosurg. Clin. N. Am. 25, 629-638 (2014).
- 13. Willis, W. D. Nociceptive Pathways: Anatomy and Physiology of Nociceptive Ascending Pathways. Philos. Trans. R. Soc. B Biol. Sci. 308, 253-268 (1985).
- 14. Treede, R. D., Kenshalo, D. R., Gracely, R. H. & Jones, A. K. P. The cortical representation of pain. Pain 79, 105-111 (1999).
- 15. Treede, R. D. Spinothalamic and thalamocortical nociceptive pathways. J. Pain 3, 109-114 (2002).
- 16. Li, J. N. & Sheets, P. L. The central amygdala to periaqueductal gray pathway comprises intrinsically distinct neurons differentially affected in a model of inflammatory pain. *J. Physiol.* 596, 6289–6305 (2018).
- 17. Veinante, P., Yalcin, I. & Barrot, M. The amygdala between sensation and affect: a role in pain. J. Mol. psychiatry $1, 9$ (2013).
- 18. Neugebauer, V. in *Mol. Pain* 265–280 (Springer New York, 2007).
- 19. Ong, W. Y., Stohler, C. S. & Herr, D. R. Role of the Prefrontal Cortex in Pain Processing. *Mol.* Neurobiol. 1–30 (2018).
- 20. Lemke, K. A. Understanding the pathopysiology of perioperative pain. Can. Vet. J. 45, 405–413 (2004).
- 21. Braz, J., Solorzano, C., Wang, X. & Basbaum, A. I. Transmitting pain and itch messages: a contemporary view of the spinal cord circuits that generate gate control. Neuron 82, 522-536 (2014).
- 22. Lima, D. & Almeida, A. The medullary dorsal reticular nucleus as a pronociceptive centre of the pain control system. Prog. Neurobiol. 66, 81-108 (2002).
- 23. Tavares, I. & Lima, D. The caudal ventrolateral medulla as an important inhibitory modulator of pain transmission in the spinal cord. J. Pain 3, 337-346 (2002).
- 24. Pinto-Ribeiro, F., Ansah, O. B., Almeida, A. & Pertovaara, A. Influence of arthritis on descending modulation of nociception from the paraventricular nucleus of the hypothalamus. Brain Res. 1197, 63–75 (2008).
- 25. Zhang, F., Vadakkan, K. I., Kim, S. S., Wu, L. J., Shang, Y. & Zhuo, M. Selective activation of microglia in spinal cord but not higher cortical regions following nerve injury in adult mouse. Mol. Pain **4,** 15 (2008).
- 26. Heinricher, M. M., Tavares, I., Leith, J. L. & Lumb, B. M. Descending control of nociception: Specificity, recruitment and plasticity. Brain Res. Rev. 60, 214–225 (2009).
- 27. Ren, K. & Dubner, R. Enhanced descending modulation of nociception in rats with persistent hindpaw inflammation. J. Neurophysiol. 76, 3025-3037 (1996).
- 28. Terayama, R., Guan, Y., Dubner, R. & Ren, K. Activity-induced plasticity in brain stem pain modulatory circuitry after inflammation. Neuroreport 11, 1915–1919 (2000).
- 29. Gauriau, C. & Bernard, J. Pain pathways and parabrachial circuits in the rat. Exp. Physiol. 87, 251–258 (2001).
- 30. Tracey, I. & Mantyh, P. W. The Cerebral Signature for Pain Perception and Its Modulation. Neuron 55, 377–391 (2007).
- 31. Fields, H. L., Bry, J., Hentall, I., Zorman, G., Vanderah, T., Lai, J. & Porreca, F. The activity of neurons in the rostral medulla of the rat during withdrawal from noxious heat. J. Neurosci. 3, 2545–2452 (1983).
- 32. Heinricher, M. M. & Kaplan, H. J. GABA-mediated inhibition in rostral ventromedial medulla: role in nociceptive modulation in the lightly anesthetized rat. Pain 47 , 105–113 (1991).
- 33. Heinricher, M. M. & Tortorici, V. Interference with GABA transmission in the rostral ventromedial medulla: Disinhibition of off-cells as a central mechanism in nociceptive modulation. Neuroscience 63, 533–546 (1994).
- 34. Heinricher, M. M. & Ingram, S. L. The Brainstem and Nociceptive Modulation. Senses A Compr. Ref. 5, 593–626 (2010).
- 35. Meng, I. D., Johansen, J. P., Harasawa, I. & Fields, H. L. Kappa Opioids Inhibit Physiologically Identified Medullary Pain Modulating Neurons and Reduce Morphine Antinociception. J. Neurophysiol. 93, 1138–1144 (2004).
- 36. Heinricher, M. M., McGaraughty, S. & Tortorici, V. Circuitry Underlying Antiopioid Actions of Cholecystokinin Within the Rostral Ventromedial Medulla. J. Neurophysiol. 85, 280-286 (2017).
- 37. Heinricher, M. M. & Neubert, M. J. Neural Basis for the Hyperalgesic Action of Cholecystokinin in the Rostral Ventromedial Medulla. J. Neurophysiol. **92,** 1982–1989 (2004).
- 38. Neubert, M. J., Kincaid, W. & Heinricher, M. M. Nociceptive facilitating neurons in the rostral ventromedial medulla. Pain 110, 158-165 (2004).
- 39. Wall, P. D. The laminar organization of dorsal horn and effects of descending impulses. J. Physiol. 188, 403–423 (1967).
- 40. Sorkin, L. S., McAdoo, D. J. & Willis, W. D. Raphe magnus stimulation-induced antinociception in the cat is associated with release of amino acids as well as serotonin in the lumbar dorsal horn. Brain Res. 618, 95-108 (1993).
- 41. Martin, R. F., Haber, L. H. & Willis, W. D. Primary afferent depolarization of identified cutaneous fibers following stimulation in medial brain stem. J. Neurophysiol. **42,** 779–790 (1979).
- 42. Cui, J., Connor, W. T. O., Ungerstedt, U. & Linderoth, B. Spinal cord stimulation attenuates augmented dorsal horn release of excitatory amino acids in mononeuropathy via a GABAergic mechanism. Pain 73, 87-95 (1997).
- 43. Wilcox, T. K., Yezierski, R. P., Willis, W. D., Giesler, G. J. & Gerhart, K. D. Postsynaptic inhibition of primate spinothalamic neurons by stimulation in nucleus raphe magnus. *Brain Res.* **204.** 184– 188 (2003).
- 44. Westlund, K. N., Carlton, S. M., Zhang, D. & Willis, W. D. Direct catecholaminergic innervation of primate spinothalamic tract neurons. J. Comp. Neurol. 299, 178-186 (1990).
- 45. Tattersall, J. E., Cervero, F. & Lumb, B. M. Effects of reversible spinalization on the visceral input to viscerosomatic neurons in the lower thoracic spinal cord of the cat. J. Neurophysiol. 56, 785– 796 (2017).
- 46. Haber, L. H., Martin, R. F., Chung, J. M. & Willis, W. D. Inhibition and excitation of primate spinothalamic tract neurons by stimulation in region of nucleus reticularis gigantocellularis. J. Neurophysiol. **43,** 1578–1593 (2017).
- 47. Treede, R.-D., Rief, W., Barke, A., Aziz, Q., Bennett, M. I., Benoliel, R., Cohen, M., Evers, S., Finnerup, N. B., First, M. B., Giamberardino, M. A., Kaasa, S., Kosek, E., Lavand'homme, P., Nicholas, M., Perrot, S., Scholz, J., Schug, S., Smith, B. H., Svensson, P., Vlaeyen, J. W. S. & Wang, S.-J. A classification of chronic pain for ICD-11. Pain 156, 1 (2015).
- 48. Goldberg, D. S. & McGee, S. J. Pain as a global public health priority. *BMC Public Health* 11, 770 (2011).
- 49. Breivik, H., Collett, B., Ventafridda, V., Cohen, R. & Gallacher, D. Survey of chronic pain in Europe: Prevalence, impact on daily life, and treatment. Eur. J. Pain 10, 287–333 (2006).
- 50. Woolf, C. J. Central sensitization: Implications for the diagnosis and treatment of pain. Pain 152. 1–31 (2011).
- 51. Latremoliere, A. & Woolf, C. Central Sensitization: a generator of pain hypersensitivity by Central Neural Plasticity. J Pain 10, 895-926 (2010).
- 52. Jensen, T. S., Baron, R., Haanpää, M., Kalso, E., Loeser, J. D., Rice, A. S. C. & Treede, R. D. A new definition of neuropathic pain. Pain 152, 2204-2205 (2011).
- 53. Baron, R. Mechanisms of disease: Neuropathic pain A clinical perspective. Nat. Clin. Pract. Neurol. **2**, 95-106 (2006).
- 54. Colloca, L., Ludman, T., Bouhassira, D., Baron, R., Dickenson, A. H., Yarnitsky, D., Freeman, R., Truini, A., Attal, N., Finnerup, N. B., Eccleston, C., Kalso, E., Bennett, D. L., Dworkin, R. H. & Raja, S. N. Neuropathic pain. Nat. Rev. Dis. Prim. 3, 17002 (2017).
- 55. Bouhassira, D., Lantéri-Minet, M., Attal, N., Laurent, B. & Touboul, C. Prevalence of chronic pain with neuropathic characteristics in the general population. Pain **136,** 380–387 (2008).
- 56. Lumley, M. A., Cohen, J. L., Borszcz, G. S., Cano, A., Radcliffe, A. M., Porter, L. S., Schubiner, H. & Keefe, F. J. Pain and emotion: A biopsychosocial review of recent research. J. Clin. Psychol. 67, 942–968 (2011).
- 57. Finnerup, N. B., Sindrup, S. H. & Jensen, T. S. The evidence for pharmacological treatment of neuropathic pain. *Pain* 150, 573–581 (2010).
- 58. Taylor, R. S. Epidemiology of Refractory Neuropathic Pain. Pain Pract. 6, 22–26 (2006).
- 59. Rowbotham, D. J. Neuropathic pain and quality of life. *Eur. J. Pain* 6, 19–24 (2002).
- 60. Wittchen, H. U., Jacobi, F., Rehm, J., Gustavsson, A., Svensson, M., Jönsson, B., Olesen, J., Allgulander, C., Alonso, J., Faravelli, C., Fratiglioni, L., Jennum, P., Lieb, R., Maercker, A., van Os, J., Preisig, M., Salvador-Carulla, L., Simon, R. & Steinhausen, H. C. The size and burden of mental disorders and other disorders of the brain in Europe 2010. *Eur. Neuropsychopharmacol.* 21, 655–679 (2011).
- 61. Yalcin, I., Barthas, F. & Barrot, M. Emotional consequences of neuropathic pain: Insight from preclinical studies. Neurosci. Biobehav. Rev. 47, 154-164 (2014).
- 62. Radat, F., Margot-Duclot, A. & Attal, N. Psychiatric co-morbidities in patients with chronic peripheral neuropathic pain: A multicentre cohort study. *Eur. J. Pain (United Kingdom)* 17, 1547– 1557 (2013).
- 63. Peters, M. L. Emotional and cognitive influences on pain experience. Mod. Trends Pharmacopsychiatry 30, 138-152 (2015).
- 64. Roelofs, J., Peters, M. L., Deutz, J., Spijker, C. & Vlaeyen, J. W. S. The Fear of Pain Questionnaire (FPQ): Further psychometric examination in a non-clinical sample. Pain 116, 339–346 (2005).
- 65. Robinson, M. E., Bialosky, J. E., Bishop, M. D., Price, D. D. & George, S. Z. Supra-threshold scaling, temporal summation, and after-sensation: Relationships to each other and anxiety/fear. J. Pain Res. 3, 25-32 (2010).
- 66. Hirsh, A. T., George, S. Z., Bialosky, J. E. & Robinson, M. E. Fear of Pain, Pain Catastrophizing, and Acute Pain Perception: Relative Prediction and Timing of Assessment. J. Pain 9, 806-812 (2009).
- 67. Rhudy, J. L. & Meagher, M. W. Fear and anxiety: Divergent effects on human pain thresholds. Pain 84, 65-75 (2000).
- 68. Theunissen, M., Peters, M. L., Bruce, J., Gramke, H. F. & Marcus, M. A. Preoperative anxiety and catastrophizing: A systematic review and meta-analysis of the association with chronic postsurgical pain. *Clin. J. Pain* 28, 819-841 (2012).
- 69. Zale, E. L., Lange, K. L., Fields, S. A. & Ditre, J. W. The relation between pain-related fear and disability: a meta-analysis. *J. Pain* 14, 1019-1030 (2013).
- 70. Kehlet, H., Jensen, T. S. & Woolf, C. J. Persistent postsurgical pain: risk factors and prevention. Lancet 367, 1618-1625 (2006).
- 71. Wiech, K., Ploner, M., Lee, M. C., Bingel, U. & Tracey, I. Prestimulus functional connectivity determines pain perception in humans. Proc. Natl. Acad. Sci. 107, 355-360 (2009).
- 72. Roy, M., Piché, M., Chen, J.-I., Peretz, I. & Rainville, P. Cerebral and spinal modulation of pain by emotions. Proc. Natl. Acad. Sci. U. S. A. 106, 20900-20905 (2009).
- 73. Jennings, E. M., Okine, B. N., Roche, M. & Finn, D. P. Stress-induced hyperalgesia. Prog. Neurobiol. 121, 1–18 (2014).
- 74. Maletic, V. & Raison, C. L. Neurobiology of depression, fibromyalgia and neuropathic pain. Front. Biosci. (Landmark Ed. 14, 5291 (2009).
- 75. Bair, M. J., Robinson, R. L., Katon, W. & Kroenke, K. Depression and Pain Comorbidity: A Literature Review. Arch. Intern. Med. 163, 2433-2445 (2003).
- 76. Gaskin, M. E., Greene, A. F., Robinson, M. E. & Geisser, M. E. Negative affect and the experience of chronic pain. *J. Psychosom. Res.* **36.** 707–713 (1992).
- 77. Giesecke, T., Gracely, R. H., Williams, D. A., Geisser, M. E., Petzke, F. W. & Clauw, D. J. The relationship between depression, clinical pain, and experimental pain in a chronic pain cohort. Arthritis Rheum. **52,** 1577-1584 (2005).
- 78. Schweinhardt, P., Kalk, N., Wartolowska, K., Chessell, I., Wordsworth, P. & Tracey, I. Investigation into the neural correlates of emotional augmentation of clinical pain. Neuroimage 40, 759-766 (2008).
- 79. Moriarty, O., McGuire, B. E. & Finn, D. P. The effect of pain on cognitive function: A review of clinical and preclinical research. Prog. Neurobiol. 93, 385-404 (2011).
- 80. Bushnell, M. C., Čeko, M. & Low, L. A. Cognitive and emotional control of pain and its disruption in chronic pain. Nat. Rev. Neurosci. **14,** 502–511 (2013).
- 81. Defrin, R., Amanzio, M., De Tommaso, M., Dimova, V., Filipovic, S., Finn, D. P., Gimenez-Llort, L., Invitto, S., Jensen-Dahm, C., Lautenbacher, S., Oosterman, J. M., Petrini, L., Pick, C. G., Pickering, G., Vase, L. & Kunz, M. Experimental pain processing in individuals with cognitive impairment: Current state of the science. Pain 156, 1396-1408 (2015).
- 82. Villemure, C. & Schweinhardt, P. Supraspinal pain processing: Distinct roles of emotion and attention. Neuroscientist **16,** 276-284 (2010).
- 83. Van Damme, S., Legrain, V., Vogt, J. & Crombez, G. Keeping pain in mind: A motivational account of attention to pain. Neurosci. Biobehav. Rev. 34, 204-213 (2010).
- 84. Colloca, L., Sigaudo, M. & Benedetti, F. The role of learning in nocebo and placebo effects. Pain 136, 211–218 (2008).
- 85. Benedetti, F. Mechanisms of Placebo and Placebo-Related Effects Across Diseases and Treatments. Annu. Rev. Pharmacol. Toxicol. 48, 33-60 (2007).
- 86. Benedetti, F., Lanotte, M., Lopiano, L. & Colloca, L. When words are painful: Unraveling the mechanisms of the nocebo effect. Neuroscience 147, 260-271 (2007).
- 87. Lee, M. C., Wiech, K., Wanigasekera, V., Tracey, I., Ploner, M., Bingel, U. & Ni Mhuircheartaigh, R. The Effect of Treatment Expectation on Drug Efficacy: Imaging the Analgesic Benefit of the Opioid Remifentanil. Sci. Transl. Med. 3, 70ra14 (2011).
- 88. Cormier, S., Piché, M. & Rainville, P. Expectations modulate heterotopic noxious counterstimulation analgesia. *J. Pain* 14, 114–125 (2013).
- 89. Colleoni, M. & Sacerdote, P. Murine models of human neuropathic pain. Biochim. Biophys. Acta 1802, 924–33 (2010).
- 90. Jaggi, A. S., Jain, V. & Singh, N. Animal models of neuropathic pain. *Fundam. Clin. Pharmacol.* 25, 1–28 (2011).
- 91. Bennett, G. J. & Xie, Y. K. A peripheral mononeuropathy in rat that produces disorders of pain sensation like those seen in man. Pain 33, 87-107 (1988).
- 92. Decosterd, I. & Woolf, C. J. Spared nerve injury: an animal model of persistent peripheral neuropathic pain. Pain 87, 149-158 (2000).
- 93. Kim, S. H., Chung, J. M., Ho Kim, S. & Mo Chung, J. An experimental model for peripheral neuropathy produced by segmental spinal nerve ligation in the rat. Pain 50, 355–363 (1992).
- 94. Wasserman, J. K. & Koeberle, P. D. Development and characterization of a hemorrhagic rat model of central post-stroke pain. Neuroscience 161, 173-183 (2009).
- 95. Marques, S. A., Garcez, V. F., Del Bel, E. A. & Martinez, A. M. B. A simple, inexpensive and easily reproducible model of spinal cord injury in mice: Morphological and functional assessment. J. Neurosci. Methods 177, 183-193 (2009).
- 96. Yezierski, R. P. & Park, S. H. The mechanosensitivity of spinal sensory neurons following intraspinal injections of quisqualic acid in the rat. *Neurosci. Lett.* **157,** 115–119 (1993).
- 97. Watson, B. D., Prado, R., Dalton Dietrich, W., Ginsberg, M. D. & Green, B. A. Photochemically induced spinal cord injury in the rat. Brain Res. 367, 296-300 (1986).
- 98. Christensen, M. D., Everhart, A. W., Pickelman, J. T. & Hulsebosch, C. E. Mechanical and thermal allodynia in chronic central pain following spinal cord injury. Pain 68, $97-107$ (1996).
- 99. Kim, J., Yoon, Y. W., Hong, S. K. & Na, H. S. Cold and mechanical allodynia in both hindpaws and tail following thoracic spinal cord hemisection in rats: time courses and their correlates. Neurosci. Lett. **343,** 200–204 (2003).
- 100. Wall, P. D., Devor, M., Inbal, R., Scadding, J. W., Schonfeld, D., Seltzer, Z. & Tomkiewicz, M. M. Autotomy following peripheral nerve lesions: experimental anesthesia dolorosa. Pain 7, 103-113 (1979).
- 101. Wall, P. D., Scadding, J. W. & Tomkiewicz, M. M. The production and prevention of experimental anesthesia dolorosa. Pain 6, 175-182 (1979).
- 102. Carlton, S. M., Lekan, H. A., Kim, S. H. & Chung, J. M. Behavioral manifestations of an experimental model for peripheral neuropathy produced by spinal nerve ligation in the primate. Pain 56, 155-166 (1994).
- 103. Kiso, T., Watabiki, T., Tsukamoto, M., Okabe, M., Kagami, M., Nishimura, K., Aoki, T. & Matsuoka, N. Pharmacological characterization and gene expression profiling of an L5/L6 spinal nerve ligation model for neuropathic pain in mice. Neuroscience 153, 492-500 (2008).
- 104. Bourquin, A. F., Süveges, M., Pertin, M., Gilliard, N., Sardy, S., Davison, A. C., Spahn, D. R. & Decosterd, I. Assessment and analysis of mechanical allodynia-like behavior induced by spared nerve injury (SNI) in the mouse. Pain 122, 14e1-14e14 (2006).
- 105. Shields, S. D., Eckert, W. A. & Basbaum, A. I. Spared nerve injury model of neuropathic pain in the mouse: a behavioral and anatomic analysis. *J. Pain* 4, 465–470 (2003).
- 106. Rodrigues-Filho, R., Santos, A. R. ., Bertelli, J. A. & Calixto, J. B. Avulsion injury of the rat brachial plexus triggers hyperalgesia and allodynia in the hindpaws: a new model for the study of neuropathic pain. Brain Res. **982,** 186-194 (2003).
- 107. Quintão, N. L. M., Balz, D., Santos, A. R. S., Campos, M. M. & Calixto, J. B. Long-lasting neuropathic pain induced by brachial plexus injury in mice: Role triggered by the pro-inflammatory cytokine, tumour necrosis factor α . Neuropharmacology **50,** 614–620 (2006).
- 108. Gazelius, B., Cui, J. G., Svensson, M., Meyerson, B. & Linderoth, B. Photochemically induced ischaemic lesion of the rat sciatic nerve. A novel method providing high incidence of mononeuropathy. Neuroreport 7, 2619-2623 (1996).
- 109. Hao, J. X., Blakeman, K. H., Yu, W., Hultenby, K., Xu, X. J. & Wiesenfeld-Hallin, Z. Development of a mouse model of neuropathic pain following photochemically induced ischemia in the sciatic nerve. Exp. Neurol. 163, 231-238 (2000).
- 110. Mosconi, T. & Kruger, L. Fixed-diameter polyethylene cuffs applied to the rat sciatic nerve induce a painful neuropathy: Ultrastructural morphometric analysis of axonal alterations. Pain 64, 37-57 (1996).
- 111. Benbouzid, M., Pallage, V., Rajalu, M., Waltisperger, E., Doridot, S., Poisbeau, P., Freund-Mercier, M. J. & Barrot, M. Sciatic nerve cuffing in mice: A model of sustained neuropathic pain. *Eur. J.* Pain 12, 591-599 (2008).
- 112. Back, S. K., Sung, B., Hong, S. K. & Na, H. S. A mouse model for peripheral neuropathy produced by a partial injury of the nerve supplying the tail. *Neurosci. Lett.* 322, 153–156 (2002).
- 113. Hulse, R., Wynick, D. & Donaldson, L. F. Characterization of a novel neuropathic pain model in mice. Neuroreport 19, 825-829 (2008).
- 114. Walczak, J. S., Pichette, V., Leblond, F., Desbiens, K. & Beaulieu, P. Behavioral, pharmacological and molecular characterization of the saphenous nerve partial ligation: A new model of neuropathic pain. Neuroscience 132, 1093-1102 (2005).
- 115. Xu, M., Aita, M. & Chavkin, C. Partial infraorbital nerve ligation as a model of trigeminal nerve injury in the mouse: behavioral, neural, and glial reactions. *J. Pain* **9,** 1036–1048 (2008).
- 116. Ahn, D. K., Lim, E. J., Kim, B. C., Yang, G. Y., Lee, M. K., Ju, J. S., Han, S. R. & Bae, Y. C. Compression of the trigeminal ganglion produces prolonged nociceptive behavior in rats. *Eur. J.* Pain 13, 568-575 (2009).
- 117. Wagner, R., DeLeo, J. A., Coombs, D. W., Willenbring, S. & Fromm, C. Spinal dynorphin immunoreactivity increases bilaterally in a neuropathic pain model. Brain Res. 629, 323-326 (1993).
- 118. DeLeo, J. A., Coombs, D. W., Willenbring, S., Colburn, R. W., Fromm, C., Wagner, R. & Twitchcll, B. B. Characterization of a neuropathic pain model: sciatic cryoneurolysis in the rat. Pain 56, 9– 16 (1994).
- 119. Na, H. S., Han, J. S., Ko, K. H. & Hong, S. K. A behavioral model for peripheral neuropathy produced in rat's tail by inferior caudal trunk injury. Neurosci. Lett. 177, 50-52 (1994).
- 120. Sung, B., Loh, H. H. & Wei, L. N. Association of kappa opioid receptor mRNA upregulation in dorsal root ganglia with mechanical allodynia in mice following nerve injury. Neurosci. Lett. 291, 163–166 (2000).
- 121. Wang, H., Bloom, O., Zhang, M., Vishnubhakat, J. M., Ombrellino, M., Che, J., Frazier, A., Yang, H., Ivanova, S., Borovikova, L., Manogue, K. R., Faist, E., Abraham, E., Andersson, J., Andersson, U., Molina, P. E., Abumrad, N. N., Sama, A. & Tracey, K. J. HMG-1 as a late mediator of endotoxin lethality in mice. Science 285, 248-251 (1999).
- 122. Chacur, M., Milligan, E. D., Gazda, L. S., Armstrong, C., Wang, H., Tracey, K. J., Maier, S. F. & Watkins, L. R. A new model of sciatic inflammatory neuritis (SIN): Induction of unilateral and bilateral mechanical allodynia following acute unilateral peri-sciatic immune activation in rats. Pain 94, 231–244 (2001).
- 123. Myers, R. R., James, H. E. & Powell, H. C. Laser injury of peripheral nerve: a model for focal endoneurial damage. J. Neurol. Neurosurg. Psychiatry 48, 1265-1268 (1985).
- 124. Aicher, S. A., Silverman, M. B., Winkler, C. W. & Bebo, B. F. Hyperalgesia in an animal model of multiple sclerosis. Pain 110, 560-570 (2004).
- 125. Olechowski, C. J., Truong, J. J. & Kerr, B. J. Neuropathic pain behaviours in a chronic-relapsing model of experimental autoimmune encephalomyelitis (EAE). Pain 141, 156-164 (2009).
- 126. Lynch, J. L., Gallus, N. J., Ericson, M. E. & Beitz, A. J. Analysis of nociception, sex and peripheral nerve innervation in the TMEV animal model of multiple sclerosis. Pain 136, 293–304 (2008).
- 127. Hasnie, F. S., Breuer, J., Parker, S., Wallace, V., Blackbeard, J., Lever, I., Kinchington, P. R., Dickenson, A. H., Pheby, T. & Rice, A. S. C. Further characterization of a rat model of varicella zoster virus-associated pain: Relationship between mechanical hypersensitivity and anxiety-related behavior, and the influence of analgesic drugs. Neuroscience 144, 1495-1508 (2007).
- 128. Fleetwood-Walker, S. M., Quinn, J. P., Wallace, C., Blackburn-Munro, G., Kelly, B. G., Fiskerstrand, C. E., Nash, A. A. & Dalziel, R. G. Behavioural changes in the rat following infection with varicellazoster virus. J. Gen. Virol. 80, 2433-2436 (1999).
- 129. Keswani, S. C., Jack, C., Zhou, C. & Hoke, A. Establishment of a Rodent Model of HIV-Associated Sensory Neuropathy. J. Neurosci. 26, 10299-10304 (2006).
- 130. Wallace, V. C. J., Blackbeard, J., Pheby, T., Segerdahl, A. R., Davies, M., Hasnie, F., Hall, S., McMahon, S. B. & Rice, A. S. C. Pharmacological, behavioural and mechanistic analysis of HIV-1 gp120 induced painful neuropathy. Pain $133, 47-63$ (2007).
- 131. Wallace, V. C. J., Blackbeard, J., Segerdahl, A. R., Hasnie, F., Pheby, T., McMahon, S. B. & Rice, A. S. C. Characterization of rodent models of HIV-gp120 and anti-retroviral- associated neuropathic pain. *Brain* 130, 2688-2702 (2007).
- 132. Drel, V. R., Mashtalir, N., Ilnytska, O., Shin, J., Li, F., Lyzogubov, V. V. & Obrosova, I. G. The leptindeficient (ob/ob) mouse: A new animal model of peripheral neuropathy of type 2 diabetes and obesity. *Diabetes* 55, 3335-3343 (2006).
- 133. Oltman, C. L., Coppey, L. J., Gellett, J. S., Davidson, E. P., Lund, D. D. & Yorek, M. A. Progression of vascular and neural dysfunction in sciatic nerves of Zucker diabetic fatty and Zucker rats. Am. J. Physiol. Metab. **289,** E113-E122 (2005).
- 134. Courteix, C., Eschalier, A. & Lavarenne, J. Streptozocin-induced diabetic rats: behavioural evidence for a model of chronic pain. $Pain$ 53, 81-88 (1993).
- 135. Shimoyama, M., Tanaka, K., Hasue, F. & Shimoyama, N. A mouse model of neuropathic cancer pain. Pain 99, 167-174 (2002).
- 136. Lee, B. H., Seong, J., Kim, U. J., Won, R. & Kim, J. Behavioral characteristics of a mouse model of cancer pain. *Yonsei Med J* 46, 252-259 (2005).
- 137. Siau, C. & Bennett, G. J. Dysregulation of cellular calcium homeostasis in chemotherapy-evoked painful peripheral neuropathy. Anesth. Analg. 102, 1485-1490 (2006).
- 138. Authier, N., Gillet, J. P., Fialip, J., Eschalier, A. & Coudore, F. A New Animal Model of Vincristine-Induced Nociceptive Peripheral Neuropathy. Neurotoxicology 24, 797-805 (2003).
- 139. Persohn, E., Canta, A., Schoepfer, S., Traebert, M., Mueller, L., Gilardini, A., Galbiati, S., Nicolini, G., Scuteri, A., Lanzani, F., Giussani, G. & Cavaletti, G. Morphological and morphometric analysis of paclitaxel and docetaxel-induced peripheral neuropathy in rats. Eur. J. Cancer 41, 1460-1466 (2005).
- 140. Flatters, S. J. L. & Bennett, G. J. Studies of peripheral sensory nerves in paclitaxel-induced painful peripheral neuropathy: Evidence for mitochondrial dysfunction. Pain 122, 245–257 (2006).
- 141. Verdú, E., Vilches, J. J., Rodríguez, F. J., Ceballos, D., Valero, A. & Navarro, X. Physiological and immunohistochemical characterization of cisplatin-induced neuropathy in mice. Muscle and Nerve 22, 329–340 (1999).
- 142. Szilvássy, J., Sziklai, I., Racz, T., Horvath, P., Rabloczky, G. & Szilvassy, Z. Impaired bronchomotor responses to field stimulation in guinea-pigs with cisplatin-induced neuropathy. Eur. J. Pharmacol. 403, 259–265 (2000).
- 143. Yamamoto, K., Tsuboi, M., Kambe, T., Abe, K., Nakatani, Y., Kawakami, K., Utsunomiya, I. & Taguchi, K. Oxaliplatin administration increases expression of the voltage-dependent calcium channel α 2δ-1 subunit in the rat spinal cord. J. Pharmacol. Sci. 130, 117–122 (2016).
- 144. Ling, B., Authier, N., Balayssac, D., Eschalier, A. & Coudore, F. Behavioral and pharmacological description of oxaliplatin-induced painful neuropathy in rat. Pain 128, 225–234 (2007).
- 145. Cavaletti, G., Tredici, G., Petruccioli, M. G., Dondè, E., Tredici, P., Marmiroli, P., Minoia, C., Ronchi, A., Bayssas, M. & Griffon Etienne, G. Effects of different schedules of oxaliplatin treatment on the peripheral nervous system of the rat. *Eur. J. Cancer* 37, 2457–2463 (2001).
- 146. Suter, U., Welcher, A. A., Özcelik, T., Snipes, G. J., Kosaras, B., Francke, U., Billings-Gagliardi, S., Sidman, R. L. & Shooter, E. M. Trembler mouse carries a point mutation in a myelin gene. *Nature* 356, 241–244 (1992).
- 147. Henry, E. W., Cowen, J. S. & Sidman, R. L. Comparison of trembler and trembler-j mouse phenotypes: Varying severity of peripheral hypomyelination. J. Neuropathol. Exp. Neurol. 42, 688–706 (1983).
- 148. Adlkofer, K., Martini, R., Aguzzi, A., Zielasek, J., Toyka, K. V. & Suter, U. Hypermyelination and demyelinating peripheral neuropathy in Pmp22-deficient mice. Nat. Genet. 11, 274–280 (1995).
- 149. Adlkofer, K., Frei, R., Neuberg, D. H., Zielasek, J., Toyka, K. V & Suter, U. Heterozygous peripheral myelin protein 22-deficient mice are affected by a progressive demyelinating tomaculous neuropathy. J. Neurosci. 17, 4662-4671 (1997).
- 150. Dina, O. A., Barletta, J., Chen, X., Mutero, A., Martin, A., Messing, R. O. & Levine, J. D. Key role for the epsilon isoform of protein kinase C in painful alcoholic neuropathy in the rat. J. Neurosci. 20, 8614–8619 (2000).
- 151. Dina, O. A., Messing, R. O. & Levine, J. D. Ethanol withdrawal induces hyperalgesia mediated by PKCe. Eur. J. Neurosci. **24,** 197-204 (2006).
- 152. Perry, T. A., Weerasuriya, A., Mouton, P. R., Holloway, H. W. & Greig, N. H. Pyridoxine-induced toxicity in rats: A stereological quantification of the sensory neuropathy. Exp. Neurol. 190, 133– 144 (2004).
- 153. Chung, J. Y., Choi, J. H., Hwang, C. Y. & Youn, H. G. Pyridoxine induced neuropathy by subcutaneous administration in dogs. J. Vet. Sci. 9, 127-131 (2008).
- 154. Roveroni, R. C., Parada, C. A., Cecília, M., Veiga, F. A. & Tambeli, C. H. Development of a behavioral model of TMJ pain in rats: The TMJ formalin test. Pain 94, 185-191 (2001).
- 155. Vahidy, W. H., Ong, W. Y., Farooqui, A. A. & Yeo, J. F. Effects of intracerebroventricular injections of free fatty acids, lysophospholipids, or platelet activating factor in a mouse model of orofacial pain. Exp. Brain Res. 174, 781-785 (2006).
- 156. Gold, B. G., Voda, J., Yu, X. & Gordon, H. The immunosuppressant FK506 elicits a neuronal heat shock response and protects against acrylamide neuropathy. Exp. Neurol. 187, 160–170 (2004).
- 157. Li, S. X., Cui, N., Zhang, C. L., Zhao, X. L., Yu, S. F. & Xie, K. Q. Effect of subchronic exposure to acrylamide induced on the expression of bcl-2, bax and caspase-3 in the rat nervous system. Toxicology 217, 46-53 (2006).
- 158. Heffernan, L. P. & Benstead, T. J. Compression and entrapment syndromes. Can. Fam. Physician 33, 681–685 (1987).
- 159. Arnold, W. D. & Elsheikh, B. H. Entrapment neuropathies. Neurol. Clin. 31, 405-424 (2013).
- 160. Maves, T. J., Pechman, P. S., Gebhart, G. F. & Meller, S. T. Possible chemical contribution from chromic gut sutures produces disorders of pain sensation like those seen in man. Pain 54, 57-69 (1993).
- 161. Attal, N., Jazat, F., Kayser, V. & Guilbaud, G. Further evidence for 'pain-related' behaviours in a model of unilateral peripheral mononeuropathy. Pain 41, 235–251 (1990).
- 162. Kupers, R. C., Nuytten, D., De Castro-Costa, M. & Gybels, J. M. A time course analysis of the changes in spontaneous and evoked behaviour in a rat model of neuropathic pain. Pain 50, 101– 111 (1992).
- 163. Challa, S. R. Surgical animal models of neuropathic pain: Pros and Cons. *Int. J. Neurosci.* 125. 170–174 (2015).
- 164. Sousa, N., Almeida, O. F. X. & Wotjak, C. T. A hitchhiker's guide to behavioral analysis in laboratory rodents. Genes, Brain Behav. 5, 5-24 (2006).
- 165. Missig, G., Mei, L., Vizzard, M. A., Braas, K. M., Waschek, J. A., Ressler, K. J., Hammack, S. E. & May, V. Parabrachial Pituitary Adenylate Cyclase-Activating Polypeptide Activation of Amygdala Endosomal Extracellular Signal–Regulated Kinase Signaling Regulates the Emotional Component of Pain. *Biol. Psychiatry* 81, 671–682 (2017).
- 166. Alba-Delgado, C., Cebada-Aleu, A., Mico, J. A. & Berrocoso, E. Comorbid anxiety-like behavior and locus coeruleus impairment in diabetic peripheral neuropathy: A comparative study with the chronic constriction injury model. Prog. Neuro-Psychopharmacology Biol. Psychiatry 71, 45-56 (2016).
- 167. Caspani, O., Reitz, M. C., Ceci, A., Kremer, A. & Treede, R. D. Tramadol reduces anxiety-related and depression-associated behaviors presumably induced by pain in the chronic constriction injury model of neuropathic pain in rats. *Pharmacol. Biochem. Behav.* 124, 290-296 (2014).
- 168. Grégoire, S., Michaud, V., Chapuy, E., Eschalier, A. & Ardid, D. Study of emotional and cognitive impairments in mononeuropathic rats: Effect of duloxetine and gabapentin. Pain 153, 1657– 1663 (2012).
- 169. Urban, R., Scherrer, G., Goulding, E. H., Tecott, L. H. & Basbaum, A. I. Behavioral indices of ongoing pain are largely unchanged in male mice with tissue or nerve injury-induced mechanical hypersensitivity. Pain 152, 990-1000 (2011).
- 170. Dellarole, A., Morton, P., Brambilla, R., Walters, W., Summers, S., Bernardes, D., Grilli, M. & Bethea, J. R. Neuropathic pain-induced depressive-like behavior and hippocampal neurogenesis and plasticity are dependent on TNFR1 signaling. *Brain. Behav. Immun.* **41,** 65–81 (2014).
- 171. Seno, M. D. J., Assis, D. V., Gouveia, F., Antunes, G. F., Kuroki, M., Oliveira, C. C., Santos, L. C. T., Pagano, R. L. & Martinez, R. C. R. The critical role of amygdala subnuclei in nociceptive and depressive-like behaviors in peripheral neuropathy. Sci. Rep. 8, 13608 (2018).
- 172. Çivi, S., Emmez, G., Dere, Ü. A., Börcek, A. Ö. & Emmez, H. Effects of quercetin on chronic constriction nerve injury in an experimental rat model. Acta Neurochir. (Wien). 158, 959-965 (2016).
- 173. Ferreira-Chamorro, P., Redondo, A., Riego, G., Leánez, S. & Pol, O. Sulforaphane inhibited the nociceptive responses, anxiety - And depressive-like behaviors associated with neuropathic pain and improved the anti-allodynic effects of morphine in mice. Front. Pharmacol. 9, (2018).
- 174. Filho, P. R. M., Vercelino, R., Cioato, S. G., Medeiros, L. F., de Oliveira, C., Scarabelot, V. L., Souza, A., Rozisky, J. R., Quevedo, A. da S., Adachi, L. N. S., Sanches, P. R. S., Fregni, F., Caumo, W. & Torres, I. L. S. Transcranial direct current stimulation (tDCS) reverts behavioral alterations and brainstem BDNF level increase induced by neuropathic pain model: Long-lasting effect. Prog. Neuro-Psychopharmacology Biol. Psychiatry 64, 44-51 (2016).
- 175. Liu, S. bing, Tian, Z., Guo, Y. yan, Zhang, N., Feng, B. & Zhao, M. gao. Activation of GPR30 attenuates chronic pain-related anxiety in ovariectomized mice. Psychoneuroendocrinology 53, 94–107 (2015).
- 176. Roeska, K., Ceci, A., Treede, R. D. & Doods, H. Effect of high trait anxiety on mechanical hypersensitivity in male rats. Neurosci. Lett. 464, 160–164 (2009).
- 177. Roeska, K., Doods, H., Arndt, K., Treede, R. D. & Ceci, A. Anxiety-like behaviour in rats with mononeuropathy is reduced by the analgesic drugs morphine and gabapentin. Pain 139, 349-357 (2008).
- 178. Shepherd, J. K., Grewal, S. S., Fletcher, A., Bill, D. J. & Dourish, C. T. Behavioural and pharmacological characterisation of the elevated 'zero-maze' as an animal model of anxiety. Psychopharmacology (Berl). 116, 56-64 (1994).
- 179. Braun, A. A., Skelton, M. R., Vorhees, C. V. & Williams, M. T. Comparison of the elevated plus and elevated zero mazes in treated and untreated male Sprague-Dawley rats: Effects of anxiolytic and anxiogenic agents. Pharmacol. Biochem. Behav. 97, 406 (2011).
- 180. Bravo, L., Alba-Delgado, C., Torres-Sanchez, S., Mico, J. A., Neto, F. L. & Berrocoso, E. Social stress exacerbates the aversion to painful experiences in rats exposed to chronic pain: The role of the locus coeruleus. *Pain* 154, 2014–2023 (2013).
- 181. Crawley, J. & Goodwin, F. K. Preliminary report of a simple animal behavior model for the anxiolytic effects of benzodiazepines. Pharmacol. Biochem. Behav. 13, 167-170 (1980).
- 182. Zhao, X., Wang, C., Cui, W. G., Ma, Q. & Zhou, W. H. Fisetin exerts antihyperalgesic effect in a mouse model of neuropathic pain: Engagement of spinal serotonergic system. Sci. Rep. 5, 9043 (2015).
- 183. Thomas, A., Burant, A., Bui, N., Graham, D., Yuva-Paylor, L. A. & Paylor, R. Marble burying reflects a repetitive and perseverative behavior more than novelty-induced anxiety. Psychopharmacology (Berl). 204, 361–373 (2009).
- 184. Wilkerson, J. L., Curry, Z. A., Kinlow, P. D., Mason, B. L., Hsu, K. L., Van Der Stelt, M., Cravatt, B. F. & Lichtman, A. H. Evaluation of different drug classes on transient sciatic nerve injurydepressed marble burying in mice. Pain 159, 1155–1165 (2018).
- 185. Fuchs, P. N. & McNabb, C. T. The place escape/avoidance paradigm: A novel method to assess nociceptive processing. J. Integr. Neurosci. 11, 61-72 (2012).
- 186. Slattery, D. A. & Cryan, J. F. Using the rat forced swim test to assess antidepressant-like activity in rodents. Nat. Protoc. **7,** 1009-1014 (2012).
- 187. Can, A., Dao, D. T., Terrillion, C. E., Piantadosi, S. C., Bhat, S. & Gould, T. D. The Tail Suspension Test. *J. Vis. Exp.* e3769 (2011).
- 188. Detke, M. J., Rickels, M. & Lucki, I. Active behaviors in the rat forced swimming test differentially produced by serotonergic and noradrenergic antidepressants. Psychopharmacology (Berl). 121, 66–72 (1995).
- 189. Porsolt, R. D., Anton, G., Blavet, N. & Jalfre, M. Behavioural despair in rats: A new model sensitive to antidepressant treatments. Eur. J. Pharmacol. 47, 379-391 (1978).
- 190. Bravo, L., Torres-Sanchez, S., Alba-Delgado, C., Mico, J. A. & Berrocoso, E. Pain exacerbates chronic mild stress-induced changes in noradrenergic transmission in rats. Eur. Neuropsychopharmacol. 24, 996–1003 (2014).
- 191. Bravo, L., Mico, J. A., Rey-Brea, R., Pérez-Nievas, B., Leza, J. C. & Berrocoso, E. Depressive-like states heighten the aversion to painful stimuli in a rat model of comorbid chronic pain and depression. Anesthesiology 117, 613-625 (2012).
- 192. Bessa, J. M., Mesquita, A. R., Oliveira, M., Pêgo, J. M., Cerqueira, J. J., Palha, J. A., Almeida, O. F. X. & Sousa, N. A trans-dimensional approach to the behavioral aspects of depression. Front. Behav. Neurosci. **3,** 1 (2009).
- 193. Ishikawa, T., Yasuda, S., Minoda, S., Ibuki, T., Fukuhara, K., Iwanaga, Y., Ariyoshi, T. & Sasaki, H. Neurotropin® Ameliorates Chronic Pain via Induction of Brain-Derived Neurotrophic Factor. Cell. Mol. Neurobiol. **35,** 231-241 (2015).
- 194. Yasuda, S., Yoshida, M., Yamagata, H., Iwanaga, Y., Suenaga, H., Ishikawa, K., Nakano, M., Okuyama, S., Furukawa, Y., Furukawa, S. & Ishikawa, T. Imipramine Ameliorates Pain-related Negative Emotion via Induction of Brain-derived Neurotrophic Factor. Cell. Mol. Neurobiol. 34, 1199–1208 (2014).
- 195. Fukuhara, K., Ishikawa, K., Yasuda, S., Kishishita, Y., Kim, H. K., Kakeda, T., Yamamoto, M., Norii, T. & Ishikawa, T. Intracerebroventricular 4-methylcatechol (4-MC) ameliorates chronic pain associated with depression-like behavior via induction of brain-derived neurotrophic factor (BDNF). Cell. Mol. Neurobiol. **32,** 971-977 (2012).
- 196. Hu, B., Doods, H., Treede, R. D. & Ceci, A. Depression-like behaviour in rats with mononeuropathy is reduced by the CB2-selective agonist GW405833. Pain 143, 206-212 (2009).
- 197. Blasco-Serra, A., González-Soler, E. M., Cervera-Ferri, A., Teruel-Martí, V. & Valverde-Navarro, A. A. A standardization of the Novelty-Suppressed Feeding Test protocol in rats. Neurosci. Lett. 658, 73–78 (2017).
- 198. Alba-Delgado, C., Llorca-Torralba, M., Mico, J. A. & Berrocoso, E. The onset of treatment with the antidepressant desipramine is critical for the emotional consequences of neuropathic pain. Pain 159, 2606–2619 (2018).
- 199. Wang, X. M., Pan, W., Xu, N., Zhou, Z. Q., Zhang, G. F. & Shen, J. C. Environmental enrichment improves long-term memory impairment and aberrant synaptic plasticity by BDNF/TrkB signaling in nerve-injured mice. Neurosci. Lett. 694, 93-98 (2019).
- 200. Llorca-Torralba, M., Mico, J. A. & Berrocoso, E. Behavioral effects of combined morphine and MK-801 administration to the locus coeruleus of a rat neuropathic pain model. Prog. Neuro-Psychopharmacology Biol. Psychiatry 84, 257-266 (2018).
- 201. Li, Q., Yue, N., Liu, S. Bin, Wang, Z. F., Mi, W. L., Jiang, J. W., Wu, G. C., Yu, J. & Wang, Y. Q. Effects of chronic electroacupuncture on depression- and anxiety-like behaviors in rats with chronic neuropathic pain. Evidence-based Complement. Altern. Med. 2014, 1-10 (2014).
- 202. Alba-Delgado, C., Llorca-Torralba, M., Horrillo, I., Ortega, J. E., Mico, J. A., Sánchez-Blázquez, P., Meana, J. J. & Berrocoso, E. Chronic pain leads to concomitant noradrenergic impairment and mood disorders. *Biol. Psychiatry* 73, 54–62 (2013).
- 203. Polo, S., Díaz, A. F., Gallardo, N., Leánez, S., Balboni, G. & Pol, O. Treatment With the Delta Opioid Agonist UFP-512 Alleviates Chronic Inflammatory and Neuropathic Pain: Mechanisms Implicated. Front. Pharmacol. **10,** (2019).
- 204. Barcelon, E. E., Cho, W.-H., Jun, S. B. & Lee, S. J. Brain Microglial Activation in Chronic Pain-Associated Affective Disorder. Front. Neurosci. 13, (2019).
- 205. Li, Y., Chen, C., Li, S. & Jiang, C. Ginsenoside Rf relieves mechanical hypersensitivity, depressionlike behavior, and inflammatory reactions in chronic constriction injury rats. *Phyther. Res.* 1-9 (2019).
- 206. Jiang, H., Ke, B., Liu, J., Ma, G., Hai, K., Gong, D., Yang, Z. & Zhou, C. Inhibition of Fatty Acid Amide Hydrolase Improves Depressive-Like Behaviors Independent of Its Peripheral Antinociceptive Effects in a Rat Model of Neuropathic Pain. Anesth. Analg. 1 (2018).
- 207. Jiang, X., Yan, Q., Liu, F., Jing, C., Ding, L., Zhang, L. & Pang, C. Chronic trans-astaxanthin treatment exerts antihyperalgesic effect and corrects co-morbid depressive like behaviors in mice with chronic pain. Neurosci. Lett. 662, 36-43 (2018).
- 208. Li, Y., Wang, Y., Xuan, C., Li, Y., Piao, L., Li, J. & Zhao, H. Role of the Lateral Habenula in Pain-Associated Depression. Front. Behav. Neurosci. 11, (2017).
- 209. Murad, H. & Ayuob, N. Co-Administration of Pioglitazone Improves Fluoxetine's Antinociceptive, Neuroprotective, and Antidepressant Effects in Chronic Constriction Injury in Rats. Pain Physician 18, 609–620 (2015).
- 210. Zhao, X., Yu, C., Wang, C., Zhang, J. F., Zhou, W. H., Cui, W. G., Ye, F. & Xu, Y. Chronic resveratrol treatment exerts antihyperalgesic effect and corrects co-morbid depressive like behaviors in mice with mononeuropathy: Involvement of serotonergic system. Neuropharmacology 85, 131-141 (2014).
- 211. Ishikawa, K., Yasuda, S., Fukuhara, K., Iwanaga, Y., Ida, Y., Ishikawa, J., Yamagata, H., Ono, M., Kakeda, T. & Ishikawa, T. 4-Methylcatechol prevents derangements of brain-derived neurotrophic factor and TrkB-related signaling in anterior cingulate cortex in chronic pain with depression-like behavior. Neuroreport 25, 226-232 (2014).
- 212. Zhu, Q., Sun, Y., Zhu, J., Fang, T., Zhang, W. & Li, J. X. Antinociceptive effects of sinomenine in a rat model of neuropathic pain. Sci. Rep. 4, 7270 (2014).
- 213. Zhao, X., Wang, C., Zhang, J. F., Liu, L., Liu, A. M., Ma, Q., Zhou, W. H. & Xu, Y. Chronic curcumin treatment normalizes depression-like behaviors in mice with mononeuropathy: Involvement of supraspinal serotonergic system and GABAA receptor. Psychopharmacology (Berl). 231, 2171– 2187 (2014).
- 214. Jesse, C. R., Wilhelm, E. A. & Nogueira, C. W. Depression-like behavior and mechanical allodynia are reduced by bis selenide treatment in mice with chronic constriction injury: A comparison with fluoxetine, amitriptyline, and bupropion. Psychopharmacology (Berl). 212, 513–522 (2010).
- 215. Andersen, M. L., Hoshino, K. & Tufik, S. Increased susceptibility to development of anhedonia in rats with chronic peripheral nerve injury: Involvement of sleep deprivation? Prog. Neuro-Psychopharmacology Biol. Psychiatry 33, 960–966 (2009).
- 216. Mogil, J. S. Sex differences in pain and pain inhibition: Multiple explanations of a controversial phenomenon. Nat. Rev. Neurosci. 13, 859-866 (2012).
- 217. Melchior, M., Poisbeau, P., Gaumond, I. & Marchand, S. Insights into the mechanisms and the emergence of sex-differences in pain. Neuroscience 338, 63-80 (2016).
- 218. Bartley, E. J. & Fillingim, R. B. Sex differences in pain: A brief review of clinical and experimental findings. Br. J. Anaesth. **111**, 52-58 (2013).
- 219. Niesters, M., Dahan, A., Kest, B., Zacny, J., Stijnen, T., Aarts, L. & Sarton, E. Do sex differences exist in opioid analgesia? A systematic review and meta-analysis of human experimental and clinical studies. Pain 151, 61-68 (2010).
- 220. Altemus, M., Sarvaiya, N. & Neill Epperson, C. Sex differences in anxiety and depression clinical perspectives. Front. Neuroendocrinol. 35, 320-330 (2014).
- 221. Tall, J. M., Stuesse, S. L., Cruce, W. L. R. & Crisp, T. Gender and the behavioral manifestations of neuropathic pain. Pharmacol. Biochem. Behav. 68, 99-104 (2001).
- 222. Keeley, R. J., Bye, C., Trow, J. & McDonald, R. J. Strain and sex differences in brain and behaviour of adult rats: Learning and memory, anxiety and volumetric estimates. Behav. Brain Res. 288, 118–131 (2015).
- 223. de Vries, G. J. & Södersten, P. Sex differences in the brain: The relation between structure and function. *Horm. Behav.* **55.** 589–596 (2009).
- 224. Vacca, V., Marinelli, S., Pieroni, L., Urbani, A., Luvisetto, S. & Pavone, F. Higher pain perception and lack of recovery from neuropathic pain in females: A behavioural, immunohistochemical, and proteomic investigation on sex-related differences in mice. Pain 155, 388-402 (2014).
- 225. Nicotra, L., Tuke, J., Grace, P. M., Rolan, P. E. & Hutchinson, M. R. Sex differences in mechanical allodynia: how can it be preclinically quantified and analyzed? Front. Behav. Neurosci. **8,** 40 (2014).
- 226. Mogil, J. S. & Chanda, M. L. The case for the inclusion of female subjects in basic science studies of pain. Pain 117, 1-5 (2005).
- 227. Stout Steele, M. & Bennett, R. A. Clinical Technique: Dorsal Ovariectomy in Rodents. J. Exot. Pet Med. 20. 222-226 (2011).
- 228. Chaplan, S. R., Bach, F. W., Pogrel, J. W., Chung, J. M. & Yaksh, T. L. Quantitative assessment of tactile allodynia in the rat paw. J. Neurosci. Methods 53, 55-63 (1994).
- 229. Leite-Almeida, H., Almeida-Torres, L., Mesquita, A. R., Pertovaara, A., Sousa, N., Cerqueira, J. J. & Almeida, A. The impact of age on emotional and cognitive behaviours triggered by experimental neuropathy in rats. Pain 144, 57-65 (2009).
- 230. Ottoni, E. B. EthoLog 2.2: a tool for the transcription and timing of behavior observation sessions. Behav. Res. Methods. Instrum. Comput. 32, 446-9 (2000).
- 231. Paxinos, G. & Watson, C. The Rat Brain in Stereotaxic Coordinates. Elsevier Acad. Press (Elsevier Inc., 2007).
- 232. Pinto-Ribeiro, F., Amorim, D., David-Pereira, A., Monteiro, A. M., Costa, P., Pertovaara, A. & Almeida, A. Pronociception from the dorsomedial nucleus of the hypothalamus is mediated by the rostral ventromedial medulla in healthy controls but is absent in arthritic animals. Brain Res. Bull. 99, 100–108 (2013).
- 233. Song, Z., Ansah, O. B., Meyerson, B. A., Pertovaara, A. & Linderoth, B. The rostroventromedial medulla is engaged in the effects of spinal cord stimulation in a rodent model of neuropathic pain. Neuroscience 247, 134-144 (2013).
- 234. Cora, M. C., Kooistra, L. & Travlos, G. Vaginal Cytology of the Laboratory Rat and Mouse: Review and Criteria for the Staging of the Estrous Cycle Using Stained Vaginal Smears. Toxicol. Pathol. 43, 776–793 (2015).
- 235. Van Goethem, B., Schaefers-Okkens, A. & Kirpensteijn, J. Making a rational choice between ovariectomy and ovariohysterectomy in the dog: A discussion of the benefits of either technique. Vet. Surg. 35, 136-143 (2006).
- 236. Kalra, P. S., Fawcett, C. P., Krulich, L. & McCann, S. M. The effects of gonadal steroids on plasma gonadotropins and prolactin in the rat. *Endocrinology* **92,** 1256–1268 (1973).
- 237. Ajika, K., Krulich, L., Fawcett, C. P. & McCann, S. M. Effects of estrogen on plasma and pituitary gonadotropins and prolactin, and on hypothalamic releasing and inhibiting factors. Neuroendocrinology **9**, 304-315 (1972).
- 238. Li, L. H., Wang, Z. C., Yu, J. & Zhang, Y. Q. Ovariectomy results in variable changes in nociception, mood and depression in adult female rats. PLoS One 9, e94312 (2014).
- 239. McElroy, J. F. & Wade, G. N. Short- and long-term effects of ovariectomy on food intake, body weight, carcass composition, and brown adipose tissue in rats. *Physiol. Behav.* 39, 361–365 (1987).
- 240. Deuis, J. R., Dvorakova, L. S. & Vetter, I. Methods Used to Evaluate Pain Behaviors in Rodents. Front. Mol. Neurosci. **10,** 1-17 (2017).
- 241. Cobos, E. J. & Portillo-Salido, E. 'Bedside-to-Bench' Behavioral Outcomes in Animal Models of Pain: Beyond the Evaluation of Reflexes. Curr. Neuropharmacol. **11**, 560–591 (2013).
- 242. Bonin, R. P., Bories, C. & De Koninck, Y. A simplified up-down method (SUDO) for measuring mechanical nociception in rodents using von Frey filaments. Mol. Pain 10, (2014).
- 243. Kayser, V., Desmeules, J. & Guilbaud, G. Systemic clonidine differentially modulates the abnormal reactions to mechanical and thermal stimuli in rats with peripheral mononeuropathy. Pain 60, 275–285 (1995).
- 244. Kayser, V. & Christensen, D. Antinociceptive effect of systemic gabapentin in mononeuropathic rats, depends on stimulus characteristics and level of test integration. Pain 88, 53–60 (2000).
- 245. Hao, J. X., Yu, W., Xu, X. J. & Wiesenfeld-Hallin, Z. Capsaicin-sensitive afferents mediate chronic cold, but not mechanical, allodynia-like behavior in spinally injured rats. Brain Res. 722, 177-180 (1996).
- 246. Jasmin, L., Kohan, L., Franssen, M., Janni, G. & Goff, J. R. The cold plate as a test of nociceptive behaviors: Description and application to the study of chronic neuropathic and inflammatory pain models. Pain 75, 367-382 (1998).
- 247. Vissers, K. & Meert, T. A behavioral and pharmacological validation of the acetone spray test in gerbils with a chronic constriction injury. Anesth. Analg. 101, 457-464 (2005).
- 248. Ramos, A., Pereira, E., Martins, G. C., Wehrmeister, T. D. & Izídio, G. S. Integrating the open field, elevated plus maze and light/dark box to assess different types of emotional behaviors in one single trial. Behav. Brain Res. 193, 277-288 (2008).
- 249. Cummins, R. A. & Walsh, R. N. The Open-Field Test: A Critical Review. Psychol. Bull. 83, 482-504 (1976).
- 250. Liu, M. Y., Yin, C. Y., Zhu, L. J., Zhu, X. H., Xu, C., Luo, C. X., Chen, H., Zhu, D. Y. & Zhou, Q. G. Sucrose preference test for measurement of stress-induced anhedonia in mice. Nat. Protoc. 13, 1686–1698 (2018).
- 251. Anisman, H. & Matheson, K. Stress, depression, and anhedonia: Caveats concerning animal models. Neurosci. Biobehav. Rev. 29, 525-546 (2005).
- 252. Petit-Demouliere, B., Chenu, F. & Bourin, M. Forced swimming test in mice: A review of antidepressant activity. Psychopharmacology (Berl). 177, 245-255 (2005).
- 253. Jensen, T. S. & Finnerup, N. B. Allodynia and hyperalgesia in neuropathic pain: Clinical manifestations and mechanisms. *Lancet Neurol.* 13, 924–935 (2014).
- 254. Wall, P. D., Devor, M., Inbal, R., Scadding, J. W., Schonfeld, D., Seltzer, Z. & Tomkiewicz, M. M. Autotomy following peripheral nerve lesions: experimental anesthesia dolorosa. Pain 7, 103-113 (1979).
- 255. Yalcin, I., Bohren, Y., Waltisperger, E., Sage-Ciocca, D., Yin, J. C., Freund-Mercier, M. J. & Barrot, M. A time-dependent history of mood disorders in a murine model of neuropathic pain. *Biol.* Psychiatry **70,** 946-953 (2011).
- 256. Fellah, F., Djenidi, R., Dehbi-Zebboudj, A., Frih, H. & Kerdouche, B. Effect of Sphaerococcus Coronopifolius Stackhouse 1797 on Anxiety-like Behavior induced by Sciatic Nerve Ligation in Female Wistar Rats. Int. J. Pharma Bio Sci. 9, (2018).
- 257. Puga-Olguín, A., Rodríguez-Landa, J. F., Rovirosa-Hernández, M. de J., Germán-Ponciano, L. J., Caba, M., Meza, E., Guillén-Ruiz, G. & Olmos-Vázquez, O. J. Long-term ovariectomy increases anxiety- and despair-like behaviors associated with lower Fos immunoreactivity in the lateral septal nucleus in rats. Behav. Brain Res. 360, 185-195 (2019).
- 258. Boivin, J. R., Piekarski, D. J., Wahlberg, J. K. & Wilbrecht, L. Age, sex, and gonadal hormones differently influence anxiety- and depression-related behavior during puberty in mice. Psychoneuroendocrinology 85, 78-87 (2017).
- 259. Picazo, O., Estrada-Camarena, E. & Hernandez-Aragon, A. Influence of the post-ovariectomy time frame on the experimental anxiety and the behavioural actions of some anxiolytic agents. *Eur. J.* Pharmacol. **530,** 88–94 (2006).
- 260. Lagunas, N., Calmarza-Font, I., Diz-Chaves, Y. & Garcia-Segura, L. M. Long-term ovariectomy enhances anxiety and depressive-like behaviors in mice submitted to chronic unpredictable stress. Horm. Behav. 58, 786-791 (2010).
- 261. Luo, J., Wang, T., Liang, S., Hu, X., Li, W. & Jin, F. Experimental gastritis leads to anxiety- and depression-like behaviors in female but not male rats. Behav. Brain Funct. 9, (2013).
- 262. Hacène, F., Rachedi, B. A., Fraia, A., Frih, H., Zaafour, M., Guernine, S. & Djemli, S. Gender Differences in the Prevalence of Depression (Persolt Swimming Test) in Sciatic Nerve Injury Model Wistar Rats. Glob. Vet. 14, 790-799 (2015).
- 263. Koshkina, A., Dudnichenko, T., Baranenko, D., Fedotova, J. & Drago, F. Effects of Vitamin D3 in Long-Term Ovariectomized Rats Subjected to Chronic Unpredictable Mild Stress: BDNF, NT-3, and NT-4 Implications. Nutrients 11, 1726 (2019).
- 264. Shin, M. S., Park, S. S., Lee, J. M., Kim, T. W. & Kim, Y. P. Treadmill exercise improves depressionlike symptoms by enhancing serotonergic function through upregulation of 5-HT1A expression in the olfactory bulbectomized rats. J. Exerc. Rehabil. 13, 36-42 (2017).
- 265. Mahmoud, R., Wainwright, S. R., Chaiton, J. A., Lieblich, S. E. & Galea, L. A. M. Ovarian hormones, but not fluoxetine, impart resilience within a chronic unpredictable stress model in middle-aged female rats. Neuropharmacology 107, 278-293 (2016).
- 266. de Chaves, G., Moretti, M., Castro, A. A., Dagostin, W., da Silva, G. G., Boeck, C. R., Quevedo, J. & Gavioli, E. C. Effects of long-term ovariectomy on anxiety and behavioral despair in rats. Physiol. Behav. 97, 420-425 (2009).
- 267. Marchettini, P., Lacerenza, M., Mauri, E. & Marangoni, C. Painful peripheral neuropathies. Curr. Neuropharmacol. 4, 175–181 (2006).
- 268. Carlson, J. D., Maire, J. J., Martenson, M. E. & Heinricher, M. M. Sensitization of Pain-Modulating Neurons in the Rostral Ventromedial Medulla after Peripheral Nerve Injury. J. Neurosci. 13222-13231 (2007).
- 269. Gonçalves, L., Almeida, A. & Pertovaara, A. Pronociceptive changes in response properties of rostroventromedial medullary neurons in a rat model of peripheral neuropathy. *Eur. J. Neurosci.* 26, 2188–2195 (2007).
- 270. Amorim, D., David-Pereira, A., Marques, P., Puga, S., Rebelo, P., Costa, P., Pertovaara, A., Almeida, A. & Pinto-Ribeiro, F. A role of supraspinal galanin in behavioural hyperalgesia in the rat. PLoS One **9**, e113077 (2014).
- 271. Ellrich, J., Ulucan, C. & Schnell, C. Are 'neutral cells' in the rostral ventro-medial medulla subtypes of on- and off-cells? Neurosci. Res. 38, 419-423 (2000).
- 272. Zhang, M. T., Wang, B., Jia, Y. N., Liu, N., Ma, P. S., Gong, S. S., Niu, Y., Sun, T., Li, Y. X. & Yu, J. Q. Neuroprotective effect of liquiritin against neuropathic pain induced by chronic constriction injury of the sciatic nerve in mice. *Biomed. Pharmacother*. **95,** 186–198 (2017).
- 273. Chanchal, S. K., Mahajan, U. B., Siddharth, S., Reddy, N., Goyal, S. N., Patil, P. H., Bommanahalli, B. P., Kundu, C. N., Patil, C. R. & Ojha, S. In vivo and in vitro protective effects of omeprazole against neuropathic pain. Sci. Rep. 6, (2016).
- 274. Au, A., Feher, A., McPhee, L., Jessa, A., Oh, S. & Einstein, G. Estrogens, inflammation and cognition. Front. Neuroendocrinol. 40, 87-100 (2016).