Monitoring the spreading of industrial yeast populations in the winery environment

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Introduction

adays, about 50% of the European wine production is based on ise of active dried wine yeast. These strains were selected due to good fermentation performance and to their capacity to produce ine with desirable organoleptical characteristics. From an ogical point of view, they are non-indigenous, mostly S. *visiae* strains that are annually introduced in the ecosystem junding the winery. The fate of those yeasts in the natural romment in different geographical localizations is totally own. The present study aims to evaluate the industrial starter ts' ability to survive and spread in nature, and become part of atural microflora of musts.

Materials and Methods

Samples

The wineries chosen were in close proximity to the vine, and the same industrial yeast strains have been used continuously for the last 5 years. From 6 sampling site, before and after the harvest, grapes were collected to perform small-scale fermentations (0,25-0,51). Must samples were plated when 70g/1 of CO_2 were released, and 30 randomly selected colonies were analysed.

Molecular identification

(Vinevard 1 and 2) Zymaflore VL1, Laffort Oenologie (Vineyard 1 and 2) In a first approach, the S. cerevisiae strains isolated from vineyard 1 and 2 were analysed by PCR amplification patterns of *à*-sequences [1, 2]. The strains with an identical pattern to the one obtained for VL1 were then further analysed by comparison of their mitochondrial DNA restriction patterns [3].

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K1, K34, D254, QA23, D47 (Vineyard 3) and K1, D254, Uvaline BL, BM45, AWRI2, D80 (Vineyard 4)

Uvalme BL, BM45, AWR12, D80 (Vineyard 4) In a first screen the strains isolated from vineyard 3 and 4 unable to use lysine as sole nitrogen source and unable to growth on YPD + cycloheximide (250 mg/) were selected. These strains were analyzed by Pulse Field Gel Electrophoresis (PFGE) karyotyping using the TAFE system [4].





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