Reductive biological treatment of textile effluents



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THE BEGINNING

- Collection of soil contaminated with dyes
- Growth in rich medium to select microorganisms
- Isolation of yeasts in selective medium
- Growth of yeasts in YEPD-agar medium with model azo dye
- Identification of yeasts with better decolourising activity

FACTS ABOUT AZO DYE DECOLOURISATION BY INTACT YEAST CELLS

- Reduction of the azo dye to colourless amines
- Unspecific and non-inducible activity
- Impermeant substrates
- Extracellular reduction

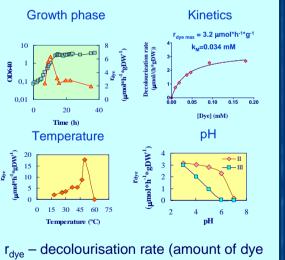


THE DECOLOURISATION PROCESS BY INTACT YEAST CELLS

10 h

5 h\

0 h



degraded per hour and per gram of biomass)

Paralel activity curves 4 동 Ę 3 ĕ 2461 2461 0. 0.01 ^ 1,2 Ferric reductase µmol*gDW⁻¹*min⁻¹) ase "min⁻¹ 0,9 0,6 ² 20 0,3 **AZO** ٥ 20 40 Time (h)

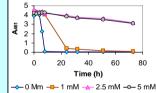
THE ENZYME SYSTEM

Since dye does not enter the cell we decided to look in the plasma membrane for redox systems.

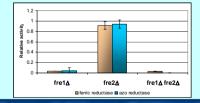
Model azo dyes used

In yeast the ferric reductase system is well characterized and reduces iron to enter the cell. Could this system be responsible for the azo reductase activity?

Iron inhibits both activities



Both activities are affected by the same magnitude by the deletion of FRE1a/o FRE2



CONCLUSIONS

- Yeasts are able to reductively decolourize azo dyes in acid effluents
- It is needed an external carbon source, that can be ethanol or glucose
- Oxygen is needed in limited conditions; under anoxic or forced aeration there is no decolourisation
- The amines produced are used as carbon and nitrogen sources the complete mineralization can be achieved
- The enzyme system responsible for the major part of this activity is the plasma membrane ferric reductase
- Dye is not toxic nor affects growth and its structure affects decolourisation
- Degradation connected to growth
- Activity detected only in intact cells